

Accuracy of current oxacillin routinely used in hospitals in Saudi Arabia

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ABSTRACT

Objective: To determine the accuracy of the current method of oxacillin resistance detection test, used in Makkah hospitals, compared with the National Committee for Clinical Laboratory Standards (NCCLS) method.

Methods: A total of 500 *S. aureus* strains were tested according to the current oxacillin sensitivity pattern used in the 4 main hospitals in Makkah between April 2003 and January 2004. In addition, the sensitivity of these clinical isolates was reconfirmed using the NCCLS standard method and polymerase chain reaction (PCR) technique.

Results: Of 500 clinical isolates, 100 (20%) were resistant to oxacillin using NCCLS standard method, but they were sensitive according to the current method.

Conclusion: A significant percent of current oxacillin resistance according to the current oxacillin sensitivity pattern used in the 4 main hospitals in Makkah between April 2003 and January 2004. In addition, the sensitivity of these clinical isolates was reconfirmed using the NCCLS standard method and PCR technique. Oxacillin resistance technique can not be used as a single test for MRSA.

Saphylococci continue to be a major cause of community-acquired bacterial infections. *Staphylococcus aureus* have acquired the ability to be coagulase-negative species of *Staphylococcus* (low-affinity penicillin-binding protein) production of PBP2a. Oxacillin-resistant (but not always) multiply resistant *S. aureus* strains are common of all antimicrobials, including macrolides, chloramphenicol, trimethoprim-sulfamethoxazole, and rifampin. The attributable increase in the use of oxacillin in hospitals is due to the fact that many are semisynthetic, penicillins, associated with carbapenems. Oxacillin-resistant strains are often (but not always) multiply resistant to several other patients.

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SCDN 1363 per admission 53. A by-product of the mecA detection of oxacillin resistance have been of a particular concern of many laboratories due to the possibility of missing some resistant strains and in the standard phenotypic tests such as the Kirby-Bauer methods due to the heterogeneity of phenotypes. A phenomenon was recognized from a study in 1994 that of ORSA; that in cultures for most strains of 15 min small proportion of cells (less than 10%) were highly resistant to oxacillin. This expressed resistance level of 10% for 30 sec is a characteristic of ORSA strains. The most common method for detection of ORSA is the detection of the gene (mecA) that encodes production of the beta-lactamase polymerase chain reaction (PCR) and chromogenic PCR. However, such methods are beyond the capabilities of many clinical laboratories (e.g. Saudi and UAE) and Administration-cleared commercial kits (e.g. EDTA performing them are not available in the Kingdom) and practical and reliable phenotypic test have been prior to of ORSA appears to be the standard of reference by Clinical Laboratory Standards Institute (CLSI) for the oxacillin-salt agar procedure (CR product).

The aim of this study was to determine the accuracy of the current ORSA detection in 100 clinical Makkan hospitals in comparison with the standard method. The PCR test was used as a standard method to compare with the current ORSA.

Methods: A total of 259 isolates with oxacillin resistance and its oxacillin sensitivity patterns were obtained from the 4 main hospitals in the Kingdom of Saudi Arabia. Oxacillin resistant strains were stored at -85°C in nutrient broth (Oxoid) containing 15% glycerol (Sigma) and were sub-cultured onto nutrient agar plates and incubated overnight at 37°C. Laboratory control in this work.

The identity of all *S. aureus* strains was confirmed following standard methods (e.g. Oxoid) on screening tests as the disc diffusion method for catalase and coagulase. Sensitivity patterns of all *S. aureus* strains were confirmed by disc diffusion method (Oxoid) using Muller Hinton agar (Oxoid) as a medium for samples under study, and the other method was Muller Hinton agar plates supplied to be more specific and incubated at 35°C for 24 hours. Accurate methods (e.g. NCCLS) any zone of inhibition of the heterogene was considered as an oxacillin sensitive strain.

Chromosomal staphylococcal DNA extracted and purified according to the method described by Pitcher et al. Primers were designed to amplify to (1) pl

Table 1 Properties of oligonucleotides primers.

Primer designation	Sequences (5-3)	Tm	Position	Amplification size (bp)
mecA(1)	AAA ATC GAT GGT AAA GGT TGG C	64.7	1828-1303	
mecA(2)	AGT TCT GCA GTA CCG GAT TTG C	662	1814-1793	533

Tm - melting temperature, bp - base pair

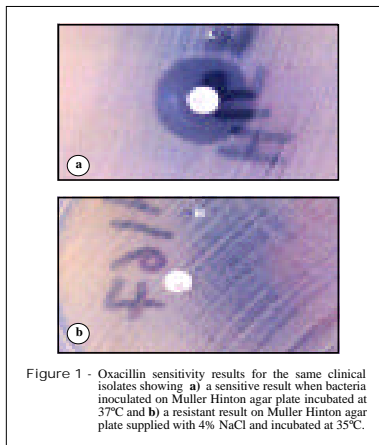


Figure 1 - Oxacillin sensitivity results for the same clinical isolates showing a) a sensitive result when bacteria inoculated on Muller Hinton agar plate incubated at 37°C and b) a resistant result on Muller Hinton agar plate supplied with 4% NaCl and incubated at 35°C.

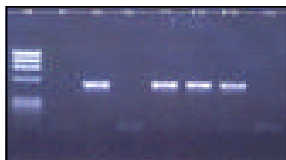


Figure 2 - Polymerase chain reaction (PCR) amplification of mecA gene demonstrating the expected 533 bp products for some of the tested samples. Lane 1 - pUC18 DNA Hae III Digest marker. Lane 2 - clinical isolate No. 51, Lane 3 - clinical isolate No. 56, Lane 4 - clinical isolate No. 76, Lane 5 - clinical isolate No. 77, Lane 6 - clinical isolate No. 471, Lane 7 - clinical isolate No. 479, Lane 8 - PCR negative (sterile water) control.

may be difficult to interpret and (ii) some isolates do not express their mecA gene unless selective pressure via antibiotic treatment is applied.¹⁰⁻¹⁷ However, in our study the PCR technique confirmed the presence of mecA gene in 88/103 samples confirmed to be MRSA using NCCLS standard technique. The discrepant findings in our study cannot be attributed to technical problems related, such as colony selection, inoculum size, or incubation time, as repeat testing yielded the same results in each of the tests. Nevertheless, these findings are in agreement with the findings of other researchers who suggested that there are other minor resistance mechanisms involved in mediating oxacillin resistance in MRSA beside the expression of mecA gene. For example, oxacillin resistance in mecA-negative strains of *S. aureus* can arise due to hyperproduction of β -lactamase, production of normal PBP with altered binding capacity, or other as unidentified factors.¹⁸ Using the PCR-based amplification technique Araj et al¹⁸ detected mecA gene in 13 out of 31 (42%) isolates initially characterized by the 1 μ g oxacillin disk diffusion test as oxacillin resistant. Unal et al¹⁹ using microdilution testing, reported that 186 of 1450 tested *S. aureus* clinical isolates were oxacillin resistant (minimal inhibitory concentration [MIC] 4 mg/ml). Fifteen of these isolates contribute conflicting results by alternative methods and were classified further. Only 2 of these (MIC 4 mg/ml) were mecA positive; 13 were inhibited by oxacillin at 4 mg/ml.

Investigators concluded that significant numbers of *S. aureus* strains classified as resistant with an oxacillin MIC of 4 mg/ml may prove susceptible by other methods. A similar finding obtained by Bignardi et al,²⁰ who evaluated several phenotypic methods for determining resistance to oxacillin. They found that, out of 44 mecA negative strains 27 were oxacillin resistant according to agar dilution test. Finally, Knapp et al²¹ noted that MRSA lacking the mecA gene could be classified as false resistant isolates by the oxacillin disk and plate methods, and attributed this to hyper-production of β -lactamase.

In conclusion, this work clearly demonstrates that a significant percentage of ORSA are currently missed diagnosed using the current sensitivity routine method which may lead to a wrong treatment choice. In addition, some mecA negative strains and oxacillin resistant can be missed diagnosed using PCR technique. This emphasizes the urgent need to comply with the recommended NCCLS guidelines.

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