

# Accuracy of current oxacillin routinely used in hospitals in Saudi Arabia

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## ABSTRACT

**Objective:** To determine the accuracy of the current method of oxacillin-resistant *Staphylococcus aureus* (ORSA) detection test, used in Makkah hospitals, compared to the method used with the National Committee for Clinical Laboratory Standards (NCCLS) method.

**Methods:** A total of 500 *S. aureus* clinical isolates and 4 main hospitals in Makkah region and Saudi Arabia, stratified between April 2003 and January 2004, were included in the study. The NCCLS standard method and a modified method using the NCCLS standard method and polymerase chain reaction (PCR) technique, oxacillin resistance technique, can not be used as a single test for ORSA.

**Results:** Of 500 clinical isolates, 100 (20%) were resistant to oxacillin using NCCLS standard method, but they were sensitive according to the modified method.

**Conclusion:** A significant percent of current clinical isolates and according to the sensitivity patterns of the modified method. In addition, the standard method and modified method can be used to determine the accuracy of the findings using the NCCLS standard method and PCR technique. Oxacillin resistance technique can not be used as a single test for ORSA.

*Saphylococci* continue to be a major cause of community-acquired bacterial infections. *Staphylococcus aureus* have acquired the ability to be oxacillin-resistant due to low-affinity penicillin-binding protein (PBP2a) production. Oxacillin-resistant strains (ORSA) per (but not always) multiply resistant to several other patients

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SCDN 1363 per admission 53. A by-product of the detection of oxacillin resistance have been of particular concern of many laboratories due to the possibility of missing some resistant strains and in the standard phenotypic tests such as the Mobic methods due to the heterogeneity of the phenomenon was recognized. Oxacillin resistance of ORSA; that in cultures for most strains of small proportion of cells (tested by the methods) were highly resistant to oxacillin. This expressed resistance level (after 1-2 hours) in susceptible strains, such as the other bacteria. Currently, the most common method for detection of ORSA is the polymerase chain reaction (PCR) that encodes production of the *meCA* gene. However, such methods are not practical and reliable for many clinical laboratories. Administration-cleared performing them are not practical and reliable. Oxacillin-salt agar procedure (CR product).

The aim of this study was to determine the accuracy of the current ORSA detection methods in Makkah hospitals in comparison with the standard method. The PCR method was used as a standard method to compare the accuracy of the ORSA.

**Methods:** A total of 259 isolates and its oxacillin sensitivity patterns were obtained from the 4 main hospitals in the Kingdom of Saudi Arabia. The isolates were stored at -85°C in nutrient broth containing 15% glycerol. They were sub-cultured onto nutrient agar and incubated overnight at 37°C. *Staphylococcus aureus* 259 control strain was used in this work.

The identity of all *S. aureus* colonies was confirmed following standard microbiological identification of Gram-positive cocci in clusters, catalase and coagulase tests. The sensitivity patterns of all *S. aureus* isolates were confirmed by disc diffusion method (Oxoid, UK) using Muller Hinton agar (Oxoid) plates. The other method used was the Hinton agar plates supplied to be more specific and incubated at 35°C for 24 hours. The results were considered as an oxacillin sensitive if there was a zone of inhibition of the colonies.

Chromosomal staphylococcal DNA extracted and purified according to the method of Pitcher et al. Primers were designed to amplify the *meCA* gene.

**Table 1** Properties of oligonucleotides primers.

Primer designation	Sequences (5-3)	Tm	Position	Amplification size (bp)
mecA(1)	AAA ATC GAT GGT AAA GGT TGG C	64.7	1828-1303	
mecA(2)	AGT TCT GCA GTA CCG GAT TTG C	662	1814-1793	533

Tm - melting temperature, bp - base pair

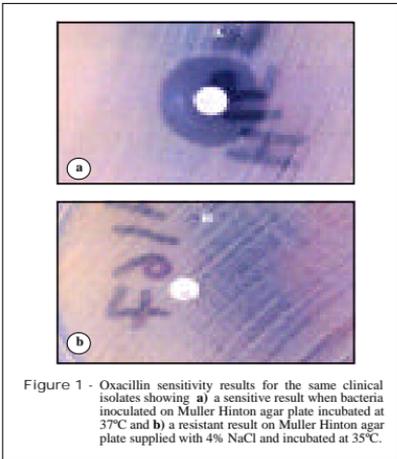


Figure 1 - Oxacillin sensitivity results for the same clinical isolates showing a) a sensitive result when bacteria inoculated on Muller Hinton agar plate incubated at 37°C and b) a resistant result on Muller Hinton agar plate supplied with 4% NaCl and incubated at 35°C.

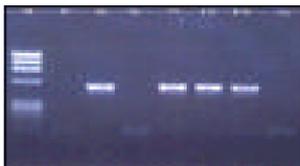


Figure 2 - Polymerase chain reaction (PCR) amplification of mecA gene demonstrating the expected 533 bp products for some of the tested samples. Lane 1 - pUC18 DNA Hae III Digest marker, Lane 2 - clinical isolate No. 51, Lane 3 - clinical isolate No. 56, Lane 4 - clinical isolate No. 76, Lane 5 - clinical isolate No. 77, Lane 6 - clinical isolate No. 471, Lane 7 - clinical isolate No. 479, Lane 8 - PCR negative (sterile water) control.

may be difficult to interpret and (ii) some isolates do not express their mecA gene unless selective pressure via antibiotic treatment is applied.<sup>10-17</sup> However, in our study the PCR technique confirmed the presence of mecA gene in 88/103 samples confirmed to be MRSA using NCCLS standard technique. The discrepant findings in our study cannot be attributed to technical problems related, such as colony selection, inoculum size, or incubation time, as repeat testing yielded the same results in each of the tests. Nevertheless, these findings are in agreement with the findings of other researchers who suggested that there are other minor resistance mechanisms involved in mediating oxacillin resistance in MRSA beside the expression of mecA gene. For example, oxacillin resistance in mecA-negative strains of *S. aureus* can arise due to hyperproduction of  $\beta$ -lactamase, production of normal PBP with altered binding capacity, or other as unidentified factors.<sup>18</sup> Using the PCR-based amplification technique Araj et al<sup>18</sup> detected mecA gene in 13 out of 31 (42%) isolates initially characterized by the 1  $\mu$ g oxacillin disk diffusion test as oxacillin resistant. Unal et al<sup>19</sup> using microdilution testing, reported that 186 of 1450 tested *S. aureus* clinical isolates were oxacillin resistant (minimal inhibitory concentration [MIC] 4 mg/ml). Fifteen of these isolates contribute conflicting results by alternative methods and were classified further. Only 2 of these (MIC 4 mg/ml) were mecA positive; 13 were inhibited by oxacillin at 4 mg/ml.

Investigators concluded that significant numbers of *S. aureus* strains classified as resistant with an oxacillin MIC of 4 mg/ml may prove susceptible by other methods. A similar finding obtained by Bignardi et al,<sup>20</sup> who evaluated several phenotypic methods for determining resistance to oxacillin. They found that, out of 44 mecA negative strains 27 were oxacillin resistant according to agar dilution test. Finally, Knapp et al<sup>21</sup> noted that MRSA lacking the mecA gene could be classified as false resistant isolates by the oxacillin disk and plate methods, and attributed this to hyper-production of  $\beta$ -lactamase.

In conclusion, this work clearly demonstrates that a significant percentage of ORSA are currently missed diagnosed using the current sensitivity routine method which may lead to a wrong treatment choice. In addition, some mecA negative strains and oxacillin resistant can be missed diagnosed using PCR technique. This emphasis the urgent need to comply with the recommended NCCLS guidelines.

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