

The role of microRNAs in regulating myocardial ischemia reperfusion injury

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ABSTRACT

يعد إفقار عضلة القلب وتضرر إعادة التروية من الحالات المرضية التي تبدأ بقصور إمداد الدم إلى القلب، ويؤدي ذلك عملية تعويض عملية التروية والتي يصاحبها الأكسجة. ويمكن أن تؤدي هذه الحالة إلى تضرر عضلة القلب، كما أنها قد تسبب العديد من الحالات المرضية بالإضافة إلى الوفاة. وبالرغم من التقدم الهائل في بحث وفهم آلية حدوث إفقار عضلة القلب وتضرر إعادة التروية إلا أن تطبيق نتائج هذا البحث في الممارسات العملية قد كان صعباً ومخيباً للآمال. يعد الحمض النووي الريبوزي المكون من فصيلة الأحماض النووية الذاتية النشوء، والأحادية الشريط، والغير مشفرة، كما أنها تندرج من فصيلة النوكليوتيد 18-24 من ناحية الطول. تعد هذه الأحماض النووية من الأحماض المحفوظة والواسعة الانتشار لدى كافة الكائنات الحية، كما ولديها القدرة على التحكم بالوظائف الخلوية المتنوعة وذلك من خلال تحسين عمليات انحلال الحمض النووي الريبوزي المكون أو منع انتقاله. ولقد أشار عدد كبير من الدراسات إلى دور هذا الحمض الفعال في العمليات الحيوية المتعلقة بأمراض القلب المزمنة والحادة. ونلخص في هذا المقال الأبحاث التي تناولت دور الحمض النووي الريبوزي المكون في عملية إفقار عضلة القلب وتضرر إعادة التروية.

Myocardial ischemia and reperfusion (I/R) injury is a pathological condition characterized by an initial restriction of blood supply to the heart followed by the subsequent restoration of perfusion and concomitant re-oxygenation. This condition may cause heart injury and contribute to morbidity and mortality. Although tremendous advances have been made in understanding the mechanisms of myocardial I/R injury, the translation of these findings into the clinical setting has been largely disappointing. MicroRNAs (miRNAs) are endogenous, single-stranded, non-coding RNAs ranging from 18-24 nucleotides in length. They are highly conserved and ubiquitously expressed in all species, which control diverse cellular functions by either promoting degradation or inhibiting target mRNA translation. In particular, a multitude of studies demonstrated miRNAs played an important role in acute and chronic cardiovascular

disease processes. In this review, we focus on miRNAs and summarize the latest insights on the role of the specific miRNAs in myocardial I/R injury.

Saudi Med J 2015; Vol. 36 (7): 787-793

doi: 10.15537/smj.2015.7.11089

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Heart damage caused by ischemia reperfusion (I/R) injury represents a serious event, which often leads to deterioration or even loss of heart function, limits the benefits of reperfusion after acute myocardial infarction (AMI) and has caused a serious global health problem.¹ Myocardial I/R injury is a complex pathophysiological event, thanks to the restoration of blood flow to the occlusion vessel.² Serious acute and chronic heart damage is aggravated because of reactive oxygen and nitrogen species and inflammatory reactions.³ Indeed, a wide range of pathological processes contribute to myocardial I/R injury (Figure 1).³ The hypoxia and the following oxidative stress result in protein modifications, lipid oxidations, and DNA breakage, triggering a chain of deleterious responses that affect all major

Disclosure. This work was supported by the National Natural Science Foundation of China (Grant No. 81200088 and 81470387), the Natural Science Foundation of Yichang City, China (Grant No. A12301-01), and Hubei Province's Outstanding Medical Academic Leader Program, Hubei, China. Authors have no conflict of interests and the work was not supported or funded by any drug company.

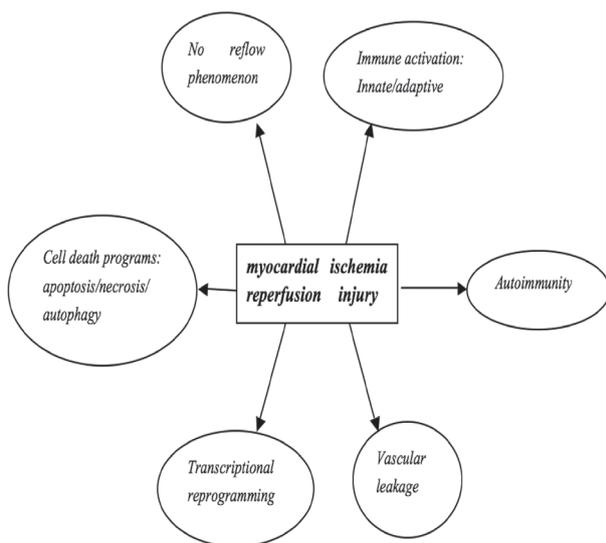


Figure 1 - Pathological processes contribute to ischemia and reperfusion associated tissue injury. Copyright permission from Eltzschig HK, Eckle T. Ischemia and reperfusion-from mechanism to translation. Nature Medicine 2011; 17: 1391-1401.

extra-and intra cellular tissue components: endothelial dysfunction, neutrophil adherence to endothelium, and trans-endothelial migration, the release of inflammatory mediators, cellular calcium overload, and eventually cell death. These events are the underlying mechanism of acute I/R damage and dysfunction in the heart. However, although tremendous advances have been made in understanding the mechanisms of myocardial I/R injury, the translation of these findings into the clinical setting has been largely disappointing.³ Despite the recent advances, the underlying molecular signaling between cellular components, extracellular matrix, and tissue vascularization associated with myocardial I/R injury are far from being completely understood. MicroRNAs have been implicated as transcriptional regulators in a wide range of biological processes determining cell fate, stress response, proliferation, or death.⁴⁻⁶ Recently, several studies have suggested that miRNAs contribute to I/R injury by altering key signaling elements; thus, making them potential therapeutic targets. Our data also has demonstrated that

Table 1 - The role of microRNAs in myocardial ischemia reperfusion injury.¹⁵

MicroRNAs	Proven targets	Biological functions
miRNA-1 ²¹	Bcl-2, HSP60, HSP70, Caspase-9, GJA1, KCNJ2	promoting apoptosis
miRNA-15 ^{21,34}	Arl2	promoting cell death
miRNA-21 ³⁶⁻⁴²	PDCD4, PTEN, FasL, Tap63 isoform of the p53 family, HNRPK, LRRFIP1 (an NF-kB inhibitor)	double-edged sword in this area
miRNA-24 ^{17,43,44}	Bcl-2	double-edged sword in this area
miRNA-29 ^{36,45,46}	Mcl-1, p85a (the regulatory subunit of PI3K), CDC42 (a Rho family GTPase), Collagen type Ia1, Collagen type Ia2, Collagen type IIIa1, Fibrillin 1, Elastin	Double-edged sword in this area
miRNA-49	Undecided	Inhibiting myocardial cell death
miRNA-17-92a ^{32,33}	Integrin subunit _5 (ITGA5)	Affecting angiogenesis
miRNA-126 ¹⁶⁻¹⁹	Spred-1, VCAM-1, CXCL12	Promoting blood vessel growth
miRNA-133 ^{20,21}	Caspase 9	inhibiting apoptosis
miRNA-144-451 ^{22,23}	CUG triplet repeat-binding protein 2	protection from myocardial injury
miRNA-145 ²⁴	CaMKIIδ,	inhibiting apoptosis
miRNA-146	Neuregulin-1 ErbB4	undecided
miRNA-149	Akt 1, E2F1	undecided
miRNA-195	Arl2	undecided
miRNA-199a ²⁵	Hif-1a, Sirt1	alteration of p53 expression/affecting cardiomyocyte apoptosis
miRNA-208	THRAP1	undecided
miRNA-210 ²⁶⁻²⁸	Caspase 8	promoting cells survival
miRNA-214 ^{22,29,30}	PTEN, NCX1, BIM, cyclophilin D	protection from cardiac cell death and hypertrophy/ fibrosis
miRNA-320 ^{21,35}	Hsp20	increasing cell death and apoptosis
miRNA-491	Bcl-X _L	undecided
miRNA-494	PTEN, FGFR2	protection from I/R injury/ apoptosis
miRNA-499 ^{22,31}	α- and β-isoforms of the calcineurin catalytic subunit SERCA2a	protection from myocardial injury
miRNA-574-3p		Undecided

HSP - heat shock protein, Arl2 - ADP ribosylation factor like 2, PDCD4 - programmed cell death protein 4, PTEN - phosphatase and tensin homolog deleted on chromosome 10, HNRPK - homo sapiens heterogeneous nuclear ribonucleo protein K, TRAP1 - involvement of tumor necrosis factor receptor-associated protein 1, VCAM-1 - vascular cell adhesion molecule 1, FGF - fibroblast growth factor, CXCL12 - CXC-motiv-chemokine 12, VEGF - vascular endothelial growth factor, CXCR4 - chemokine receptor 4, FGFR2 - fibroblast growth factor receptor 2

miRNA-22 could inhibit apoptosis of cardio myocytes through one of its targets cAmp response element binding (CREB) binding protein (CBP), which may constitute a new therapeutic target for the prevention of myocardial I/R injury.⁷ Here, we highlight the latest advances in the identification of myocardial ischemia-associated miRNAs and their functional significance in the modulation of I/R-induced cardio myocyte death (necrosis/apoptosis), myocardial inflammation, fibrosis, compromised contractile function, and neoangiogenesis (Table 1).

Source and function of microRNAs. MicroRNAs are a class of endogenous, small (~22 nucleotides [nt] in size), noncoding single-stranded RNAs.⁴ The genes are located in the intron, non-coding exon, and intergenic regions of genomes, and initially transcribed by polymerase II into primary miRNAs (pri-miRNAs).⁷ A stem-loop pre-cursor miRNA (pre-miRNA, ~70 nt in size) would be formed after the pri-miRNAs subsequently are cleaved by a microprocessor complex composed of Drosha and Pasha. Later, the formed pre-miRNAs would be transported into the cytoplasm. After this process, Dicer would cleave them into mature miRNAs.⁸ The mature miRNAs can be incorporated into the RNA-induced silencing complex (RISC) and negatively regulate gene expression through binding on the untranslated region (UTR) of target mRNAs.⁹ The miRNAs could regulate genes involved in a series of diverse biological processes, including development, differentiation, inflammation, stress response, angiogenesis, adhesion, proliferation, and apoptosis by this mechanism.^{10,11}

Expression of microRNAs in myocardial ischemia reperfusion injury. He et al¹² detected the miRNAs expression of the myocardial ischemia Sprague-Dawley (SD) rats after reperfusion in one hour, and 3 hours (I/R, 1h/3h) and found that 16 kinds of miRNAs were significantly disordered, of which 10 were up-regulated, and 6 were down-regulated. When reperfusion in 30 minutes or in 24 hours (I/R, 30min/24h), only the miRNA-1, miRNA-126, miRNA-208 were up-regulated, but miRNA-21, miRNA-133, and miRNA-195 reduced.¹³ At the same time, the myocardial ischemia FVB/N mice (offers a system suitable for transgenic experiments and subsequent genetic analyses) after reperfusion in 30 minutes or in 24 hours (I/R, 30min/24h) miRNA-320 was also observed to down-regulated, but miRNA-7, miRNA-21, miRNA-146b, and miRNA-491 increased.¹⁴ Thus, we could draw a conclusion that the miRNA are expressed in myocardial I/R depending on the dynamic nature and condition.

The miRNAs disorder expression in early reperfusion were related to myocardial cell death and oxidative stress. Therefore, analysis of the expression and function of myocardial I/R related miRNAs may be better to illustrate the mechanism of myocardial I/R injury.¹⁵

Key microRNAs, which play an important role in myocardial I/R injury. 1) **Protective microRNAs in myocardial I/R: miRNA-126, miRNA-133, miRNA-144, miRNA-145, miRNA-199, miRNA-210, miRNA-214, miRNA-494, miRNA-451, and miRNA-499.** The miRNA-126, as an angiogenic factor, is highly expressed in the heart, which could be maintained to promote blood vessel growth by enhancing the proangiogenic actions of vascular endothelial growth factor (VEGF) and fibroblast growth factor (FGF), and promoting blood vessel formation by repressing the inhibitor Spred-1 and vascular cell adhesion molecule 1 (VCAM-1).¹⁶⁻¹⁸ The expression of miRNA-126 increased after myocardial I/R. When miRNA-126 was knockout, the survival rate is significantly reduced.¹⁹ At the same time, a research by Zerneck also has proven that miRNA-126 can also inhibit apoptosis by activating the CXC-Motiv-Chemokine12 (CXCL12) chemokine receptor chemokine receptor 4 (CXCR4).¹⁹ Thus, currently we have evidence that miRNA-126 is a protective miRNAs in myocardial I/R enhancing angiogenesis and inhibiting apoptosis.

The miRNA-133 is expressed in cardio myocytes. miRNA-133 levels are reduced after myocardial I/R injury.²⁰ When researchers up-regulated the levels of miRNA-133, cell apoptosis was reduced notably and the viability of cardiomyocytes (H₉C₂) cells after exposure to hydrogen peroxide (H₂O₂) was also increased. However, after the expression of miRNA-133 was down-regulated by oligonucleotide, the results were opposite. The mechanism may be related to its targets; namely, miRNA-133 would cut the pro-apoptotic gene caspase-9 expression.²¹ Previous studies²¹ have shown that increasing miRNA-133 levels would reduce myocardial I/R injury.

The miRNA-144 and miRNA-451 were shown to protect cardio myocytes from apoptotic cell death in vitro.^{22,23} Conversely, the knockdown of endogenous miRNA-144 and miRNA-451 augmented cell death. The CUG triple repeat-binding protein2, which has been described to regulate the translation of COX-2, was identified as a bona fide target of these 2 miRNAs. The effects of miRNA-144 and miRNA-451 modulation were blocked by simultaneous inhibition of COX-2 by specific inhibitors, NS-398 or DUP-697, further high

lighting the importance of this regulatory pathway in the protection from myocardial I/R injury.^{22,23}

An increasing number of studies have declared that up-regulating the intracellular divalent calcium ions (Ca^{2+}) concentrations are closely associated with the activation of calcium/calmodulin-dependent protein kinase (CaMKII), which up-regulates the expression of apoptotic genes. Recently, Cha et al²⁴ revealed that miRNA-145 inhibited Ca^{2+} overload and Ca^{2+} -related signals by targeting CaMKII δ , and the over-expression of miRNA-145 protects against reactive oxygen species (ROS)-induced cardio myocyte apoptosis. The miRNA-199 family includes miRNA-199a-1, miRNA-199a-2, and miRNA-199b. Rane et al²⁵ reported that under anoxic conditions, the cells may rapidly reduced cardiac miRNA-199a expression by a post-transcriptional mechanism. Down-regulation miRNA-199a could induce apoptosis during hypoxia by increasing the expression of hypoxia inducible factor-1 α and sirtuin 1 (srt1). Some researchers^{24,25} also supported that knockdown miR-199a before hypoxia may play a role in pre-treatment and protection.

Just like miRNA-126, miRNA-210 has been shown to possess pro-angiogenic properties.²⁶ Up-regulation of miRNA-210 would increase the migration of vascular endothelial cells. On the contrary, when in hypoxia circumstances, decreasing the expressions of miRNA-210 would inhibit endothelial cell migration and cell survival. miRNA-210 could promote hypoxia miRNA-210-mediated cell survival, migration and angiogenesis by inhibiting ephrin-A3 gene.²⁶ Mutharasan et al²⁷ also found that miRNA-210 would be over-expressed in myocardial hypoxia factor-dependent and independent pathways, which would inhibit oxidative stress-induced apoptosis by influencing the formation of ROS. In addition, Kim et al²⁸ revealed that miRNA-210 had a function to promote cells survival by blocking caspase-8 related protein-2 after myocardial I/R. In summary, the above related studies have shown that miRNA-210 myocardial protection could be used as a new way to treat ischemic heart disease.

miRNA-214 was shown to be highly increased in the models of cardiac hypertrophy and heart failure.²² Genetic deletion of miRNA-214 aggravated myocardial I/R injury, increased apoptotic cell death, promoted the excessive accumulation of fibrotic tissue, and resulted in a loss of contractility. While, when researchers up-regulated the levels in a variety of cardiac disease models, miRNA-214 would play a protective role. Up to now, the common concept of the mechanism of this biological

function is related to its role of inhibiting the expression of the Ca^{2+} -handling molecule Na^+ - Ca^{2+} exchangers1 (NCX1), as well as CaMKII, and cyclophilin D.^{29,30}

miRNA-499 was found at intron and highly expressed in the heart.^{22,31} The expressing levels of myocardial miRNA-499 were identified as being down regulated after cardiac I/R injury. The direct targets of this miRNA were α - and β -isoforms of the calcineurin catalytic subunit. Targeting of these 2 subunits resulted in a reduction of mitochondrial dynamin-related protein-1 expression through blunted calcineurin-mediated dephosphorylation of DRP1 and subsequently an aggravation of cardio myocyte apoptosis. In addition to this, researchers have proven that the transcriptional repression of miRNA-499 during I/R injury was shown to be mediated by enhancing the expression of p53.³¹ AntagomiR knockdown miRNA-499 could also increase collagen deposition, promote myocyte hypertrophy, injury systolic function, and aggravate I/R cardiac remodeling.²² These results indicate that increasing the levels of cardiac miRNA-499 not only could reduce I/R injury, but also delay the AMI remodeling process.^{22,31}

2) Opposite effects of microRNAs in myocardial I/R: miRNA-1, miRNA-15, miRNA-92a, and miRNA-320.

miRNA-1 and miRNA-133 (discussed above) were located in the same chromosomal loci, transcribed to be mature simultaneously and independently, but with different biological functions. Studies have shown that miRNA-1 and miRNA-133 play a different role in stress-induced cardio myocyte survival, miRNA-133 inhibits apoptosis (discussed above), whereas, miRNA-1 promotes apoptosis.²¹ Increasing the miRNA-1 levels of H_2C_2 rat embryonic ventricular myocytes or rat myocardial cells could stimulate stress-induced apoptosis by down-regulating the expression of several anti-apoptotic genes such as Hsp60, Hsp70, IGF-1, and Bcl-2.²¹ In addition, injecting miRNA-1 may exacerbate cardiac arrhythmias. The above results suggest that miRNA-1 could lead to cell apoptosis by affecting anti-apoptotic genes.

The miRNA-17-92a gene cluster (encoding miRNA-17, miRNA-18a, miRNA-19a/b, miRNA-20a, and miRNA-92a) levels were increased 24 hours after coronary artery occlusion in mice. This gene cluster could affect angiogenesis dependent on cellular environment.³² Bonauer et al³³ showed that antagomir-92a improved left ventricular function, reduced myocardial infarction size, reduced apoptosis, and increased the number of new blood vessels. Researchers³³ revealed that the mechanism of this process was due to the effect of

this gene cluster on a wide of target genes, such as Srt, ITGA5, S1P1, MKK4, and eNOS. Thus, inhibiting miRNA-17-92a may potentially reduce infarct size after myocardial infarction. The miRNA-15 was also found to be dysregulated in animal models after I/R injury. When inhibiting the expression of miRNA-15 with anti-miR-15, it would protect cardio myocyte cells from hypoxia-induced death in vivo.^{21,34} The expression of miRNA-320 decreased in myocardial I/R injury.³⁵ Over-expression of cardiac-specific miRNA-320 transgenic mice increased the area of AMI and apoptosis. Antagomirs miRNA-320 reduced the myocardial infarct size. Transfection of miRNA-320 into cardiac microvascular endothelial cells inhibited angiogenesis by inhibiting IGF-1 expression.²¹ Therefore, silencing miRNA-320 expression by antagomirs could reduce myocardial cell death, and increase neovascularization.

3) Double-edged microRNAs in myocardial I/R: miRNA-21, miRNA-24, and miRNA-29. In recent years, several miRNAs have been proven to play a dual role in the pathomechanism of myocardial I/R injury. miRNA-21 was just one of these miRNAs.³⁶ miRNA-21 was found to be specifically localized to fibroblasts in the infarct region of the heart by immunohistochemistry techniques. Phosphatase and tensin homolog deleted on chromosome ten (PTEN) identified as a direct target was found to regulate the downstream expression of MMP-2.³⁷ miRNA-21 was indispensable for myocardial protection in that setting, and was found to reduce hypoxia/reoxygenation-induced cell apoptosis via regulation of its target gene programmed cell death protein 4 (PDCD4).³⁷ In addition to this, some scholars believed that miRNA-21 could inhibit some other pro-apoptotic genes involved in pro-survival pathways, such as programmed cell death protein 4 (PCDF4) and fas ligand (FasL).^{38,39} Over-expression cardiac-specific miRNA-21 transgenic mice could reduce myocardial injury,³⁸ and antagomir silence miRNA-21 could increase H₂O₂-induced myocardial necrosis and apoptosis.³⁹ Studies have shown that miRNA-21 could also promote the survival of fibroblasts and lead to myocardial fibrosis by regulating protein sprouty homolog 1 (Spry1).⁴⁰ However, another study⁴¹ found that over-expression of miRNA-21 reduced endothelial cell proliferation, migration, and angiogenesis by the regulation of ras homolog family member B (RhoB). Thus, inhibiting the expression of miRNA-21 also could promote coronary angiogenesis and restore blood flow, which plays a role in myocardial protection after myocardial infarction. In short, miRNA-21 played

a dual role in this area by influencing endothelial cell proliferation, migration, formation of new blood vessels, and cardiac cells survive through its certain targets.⁴² Like miRNA-21, miRNA-24 plays a similar role in this process.¹⁷ miRNA-24 is enriched in cardiac endothelial cells and is up-regulated after I/R injury. Blocking endothelial miRNA-24 limited the injury via prevention of endothelial apoptosis and enhancement of vascularity,⁴³ which led to preserved cardiac function and survival. However, miRNA-24 expression is down-regulated in the ischemic border after I/R. Delivery of miRNA-24 into the infarcted hearts of mice inhibited cardio myocyte apoptosis, attenuated infarct size, and reduced cardiac dysfunction.⁴⁴ This anti-apoptotic effect on cardio myocytes in vivo is partly mediated by the B-cell lymphoma (Bcl)-2 gene. Thus, miR-24 played a dual role in vascular endothelial cells and cardiac myocytes. It is crucial to intervene miR-24 at the right time. Another dual microRNA is miRNA-29. van Rooij et al⁴⁵ found that them iRNA-29 family (miRNA-29a, b, c) were down-regulated after I/R injury. miRNA-29 can regulate pro-fibrotic proteins gene expression.⁴⁶ Antagomir, (a kind of inhibitor, which can prevent miRNA and its target genes mRNA complementary pairing) silence miRNA-29 increasing the expression of collagen. In fibroblasts, over expression of miRNA-29 gene transcription could reduce collagen. In addition, miRNA-29 also negatively regulated anti-apoptotic genes such as Tcl-1, Mcl-1, YY1, p85 α , CDC42, and DNMT3.^{36,46} Thanks to its protective mechanism above, it inhibited I/R injury-induced myocardial cell death, and reduced myocardial remodeling. However, at the same time, it may also affect fibroblasts, which would suppress the treatment after myocardial I/R injury. Therefore, the expression of miRNA-29 in the heart has a double-edged effect.³⁶

In conclusion, myocardial I/R injury is determined by numerous mediators and signaling pathways including myocyte death (necrosis, apoptosis, and autophagy), myocyte hypertrophy, cardiac fibrosis, and impaired angiogenesis. In recent years, a growing number of miRNAs related to myocardial I/R injury were found. Identification of I/R-associated miRNAs is important as miRNAs targeting is becoming a topic of interest that could provide new ideas for the treatment of myocardial I/R injury. The role of different miRNAs are very different in this process. Briefly, miRNAs including miRNA-126, miRNA-133, miRNA-144, miRNA-145, miRNA-199a, miRNA-210, miRNA-214, miRNA-494, miRNA-451, miRNA-499 are protective

after myocardial I/R injury. While miRNAs including miRNA-1, miRNA-15, miRNA-92a, and miRNA-320 have the opposite effects. Complexly, some other miRNAs such as miRNA-21, miRNA-24, and miRNA-29 have a dual role. Modulation of miRNA levels directly or indirectly provides a therapeutic advantage to interfere with myocardial I/R injury, and the complexity of miRNA targets might yield different consequences in various cell types within the heart. However, up to now, the mechanism of the various specific miRNAs in myocardial I/R injury remained to be further clarified. Hence, to obtain knowledge of the potential target for the treatment of myocardial I/R injury, further study is required to ascertain the many functions of miRNAs in normal physiology and disease states.

Acknowledgment. We would like to thank Chao-Jun Yang for designing this work.

References

- Cai Y, Xu H, Yan J, Zhang L, Lu Y. Molecular targets and mechanism of action of dexmedetomidine in treatment of ischemia/reperfusion injury. *Mol Med Rep* 2014; 9: 1542-1550.
- Araszkiwicz A, Grygier M, Lesiak M, Grajek S. The impact of ischemia-reperfusion injury on the effectiveness of primary angioplasty in ST-segment elevation myocardial infarction. *Postepy Kardiologii Interwencyjnej* 2013; 9: 275-281.
- Eltzschig HK, Eckle T. Ischemia and reperfusion—from mechanism to translation. *Nat Med* 2011; 17: 1391-1401.
- Suzuki HI, Miyazono K. Emerging complexity of microRNA generation cascades. *J Biochem* 2011; 149: 15-25.
- Peterson SM, Thompson JA, Ufkin ML, Sathyanarayana P, Liaw L, Congdon CB. Common features of microRNA target prediction tools. *Front Genet* 2014; 5: 23.
- Fan ZX, Yang J. Microribonucleic acids and vascular restenosis. *Saudi Med J* 2014; 35: 796-801.
- Yang J, Chen L, Yang J, Ding J, Li S, Wu H, et al. MicroRNA-22 targeting CBP protects against myocardial ischemia-reperfusion injury through anti-apoptosis in rats. *Mol Biol Rep* 2014; 41: 555-561.
- Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J, et al. The nuclear RNase III Drosha initiates microRNA processing. *Nature* 2003; 425: 415-419.
- Newman MA, Hammond SM. Emerging paradigms of regulated microRNA Processing. *Genes Dev* 2010; 24: 1086-1092.
- Porrello ER. MicroRNAs in cardiac development and regeneration. *Clin Sci* 2013; 125: 151-166.
- Condorelli G, Latronico MV, Cavarretta E. MicroRNAs in cardiovascular diseases: Current knowledge and the road ahead. *J Am Coll Cardiol* 2014; 63: 2177-2187.
- He B, Xiao J, Ren AJ, Zhang YF, Zhang H, Chen M, et al. Role of miR-1 and miR-133a in myocardial ischemic postconditioning. *J Biomed Sci* 2011; 18: 22-31.
- Tang Y, Zheng J, Sun Y, Wu Z, Liu Z, Huang G. MicroRNA-1 regulates cardiomyocyte apoptosis by targeting Bcl-2. *Int Heart J* 2009; 50: 377-387.
- Ren XP, Wu J, Wang X, Sartor MA, Qian J, Jones K, et al. MicroRNA-320 is involved in the regulation of cardiac ischemia /reperfusion injury by targeting heat-shock protein20. *Circulation* 2009; 119: 2357-2366.
- Choi E, Cha MJ, Hwang KC. Roles of Calcium Regulating MicroRNAs in Cardiac Ischemia-Reperfusion Injury. *Cells* 2014; 3: 899-913.
- Zhang Q, Kandic I, Kutryk MJ. Dysregulation of angiogenesis-related microRNAs in endothelial progenitor cells from patients with coronary artery disease. *Biochem Biophys Res Commun* 2011; 405: 42-46.
- Zhu H, Fan GC. Role of microRNAs in the reperfused myocardium towards post-infarct remodeling. *Cardiovasc Res* 2012; 94: 284-292.
- Wang S, Aurora AB, Johnson BA, Qi X, McAnally J, Hill JA, et al. The endothelial-specific microRNA miR-126 governs vascular integrity and angiogenesis. *Dev Cell* 2008; 15: 261-271.
- Zernecke A, Bidzhekov K, Noels H, Shagdarsuren E, Gan L, Denecke B, et al. Delivery of microRNA-126 by apoptotic bodies induces CXCL12-dependent vascular protection. *Sci Signal* 2009; 2: ra81.
- Bostjancic E, Zidar N, Stajer D, Glavac D. MicroRNAs miR-1, miR-133a, miR-133b and miR-208 are dysregulated in human myocardial infarction. *Cardiology* 2010; 115: 163-169.
- Xu C, Lu Y, Pan Z, Chu W, Luo X, Lin H, et al. The muscle-specific microRNAs miR-1 and miR-133 produce opposing effects on apoptosis by targeting HSP60, HSP70 and caspase-9 in cardiomyocytes. *J Cell Sci* 2007; 120: 3045-3052.
- Lorenzen JM, Batkai S, Thum T. Regulation of cardiac and renal ischemia-reperfusion injury by microRNAs. *Free Radic Biol Med* 2013; 64: 78-84.
- Zhang X, Wang X, Zhu H, Zhu C, Wang Y, Pu WT, et al. Synergistic effects of the GATA-4-mediated miR-144/451 cluster in protection against simulated ischemia/reperfusion-induced cardio myocyte death. *J Mol Cell Cardiol* 2010; 49: 841-850.
- Cha MJ, Jang JK, Ham O, Song BW, Lee SY, Lee CY, et al. MicroRNA-145 suppresses ROS-induced Ca²⁺ overload of cardiomyocytes by targeting cAMPKII δ . *Biochem Biophys Res Commun* 2013; 435: 720-726.
- Rane S, He M, Sayed D, Vashistha H, Malhotra A, Sadoshima J, et al. Downregulation of miR-199a derepresses hypoxia-inducible factor-1 α and recapitulates hypoxia preconditioning in cardiac myocytes. *Circ Res* 2009; 104: 879-886.
- Fasanaro P, D'Alessandra Y, Di Stefano V, Melchionna R, Romani S, Pompilio G, et al. MicroRNA-210 modulates endothelial cell response to hypoxia and inhibits the receptor tyrosine kinase ligand Ephrin-A3. *J Biol Chem* 2008; 283: 15878-15883.
- Mutharasan RK, Nagpal V, Ichikawa Y, Ardehali H. MicroRNA-210 is upregulated in hypoxic cardiomyocytes through Akt- and p53-dependent pathways and exerts cytoprotective effects. *Am J Physiol Heart Circ Physiol* 2011; 301: H1519-H1530.
- Kim HW, Haider HK, Jiang S, Ashraf M. Ischemic preconditioning augments survival of stem cells via miR-210 expression by targeting caspase-8-associated protein 2. *J Biol Chem* 2009; 284: 33161-33168.
- Van Rooij E, Sutherland LB, Liu N, Williams AH, McAnally J, Gerard RD, et al. A signature pattern of stress-responsive microRNAs that can evoke cardiac hypertrophy and heart failure. *Proc Natl Acad Sci USA* 2006; 103: 18255-18260.

30. Aurora AB, Mahmoud AI, Luo X, Johnson BA, van Rooij E, Matsuzaki S, et al. MicroRNA-214 protects the mouse heart from ischemic injury by controlling Ca^{2+} overload and cell death. *J Clin Invest* 2012; 122: 1222-1232.
31. Wang JX, Jiao JQ, Li Q, Long B, Wang K, Liu JP, et al. miR-499 regulates mitochondrial dynamics by targeting calcineurin and dynamin-related protein-1. *Nat Med* 2011; 17: 71-78.
32. Doebele C, Bonauer A, Fischer A, Scholz A, Reiss Y, Urbich C, et al. Members of the microRNA-17-92 cluster exhibit a cell-intrinsic antiangiogenic function in endothelial. *Cells Blood* 2010; 115: 4944-4950.
33. Bonauer A, Carmona G, Iwasaki M, Mione M, Koyanagi M, Fischer A, et al. MicroRNA-92a controls angiogenesis and functional recovery of ischemic tissues in mice. *Science* 2009; 324: 1710-1713.
34. Hullinger TG, Montgomery RL, Seto AG, Dickinson BA, Semus HM, Lynch JM, et al. Inhibition of miR-15 protects against cardiac ischemic injury. *Circ Res* 2012; 110: 71-81.
35. Ren XP, Wu J, Wang X, Sartor MA, Qian J, Jones K, et al. MicroRNA-320 is involved in the regulation of cardiac ischemia /reperfusion injury by targeting heat-shock protein20. *Circulation* 2009; 119: 2357-2366.
36. Yumei Y, Jose R, Perez P, Qian J, Birnbaum Y. The role of microRNA in modulating myocardial ischemia-reperfusion injury. *Physiol Genomics* 2011; 43: 534-542.
37. Papagiannakopoulos T, Shapiro A, Kosik KS. MicroRNA-21 targets a network of key tumor-suppressive pathways in glioblastoma cells. *Cancer Res* 2008; 68: 8164-8172.
38. Sayed D, He M, Hong C, Gao S, Rane S, Yang Z, et al. MicroRNA-21 is a downstream effector of AKT that mediates its antiapoptotic effects via suppression of Fas ligand. *J Biol Chem* 2010; 285: 20281-20290.
39. Cheng Y, Zhu P, Yang J, Liu X, Dong S, Wang X, et al. Ischaemic preconditioning-regulated miR-21 protects heart against ischaemia /reperfusion injury via anti-apoptosis through its target PDCD4. *Cardiovasc Res* 2010; 87: 431-439.
40. Qin Y, Yu Y, Dong H, Bian X, Guo X, Dong S. MicroRNA 21 inhibits left ventricular remodeling in the early phase of rat model with ischemia-reperfusion injury by suppressing cell Apoptosis. *Int J Med Sci* 2012; 9: 413-423.
41. Sabatel C, Malvaux L, Bovy N, Deroanne C, Lambert V, Gonzalez ML, et al. MicroRNA-21 exhibits antiangiogenic function by targeting RhoB expression in endothelial cells. *PLoS One* 2011; 6: e16979.
42. Xu X, Kriegel AJ, Jiao X, Liu H, Bai X, Olson J, et al. miR-21 in Ischemia/Reperfusion Injury: A Double-edged Sword? *Physiol Genomics* 2014; 46: 789-797.
43. Fiedler J, Jazbutyte V, Kirchmaier BC, Gupta SK, Lorenzen J, Hartmann D et al. MicroRNA-24 regulates vascularity after myocardial infarction. *Circulation* 2011; 124: 720-730.
44. Qian L, Van Laake LW, Huang Y, Liu S, Wendland MF, Srivastava D. miR-24 inhibits apoptosis and represses Bim in mouse cardiomyocytes. *J Exp Med* 2011; 208: 549-560.
45. van Rooij E, Sutherland LB, Thatcher JE, DiMaio JM, Naseem RH, Marshall WS, et al. Dysregulation of microRNAs after myocardial infarction reveals a role of miR-29 in cardiac fibrosis. *Proc Natl Acad Sci USA* 2008; 105: 13027-13032.
46. Park SY, Lee JH, Ha M, Nam JW, Kim VN. miR-29 miRNAs activate p53 by targeting p85 alpha and CDC42. *Nat Struct Mol Biol* 2009; 16: 23-29.

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Zhou Q, Deng Q, Huang J, Chen JL, Hu B, Guo RQ. Evaluation of left ventricular mechanical dyssynchrony in patients with heart failure after myocardial infarction by real-time three-dimensional echocardiography. *Saudi Med J* 2012; 33: 256-261.