

Prevalence of extended-spectrum beta-lactamases among *Enterobacteriaceae* isolated from blood culture in a tertiary care hospital

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ABSTRACT

Objectives: To determine the prevalence of extended spectrum β -lactamase among *Enterobacteriaceae* isolated from blood culture in a tertiary care hospital.

Method: We carried out this study at the Armed Forces Hospital, Riyadh, Kingdom of Saudi Arabia during the period between January 2003 – December 2004. We tested a total of 601 isolates of the family *Enterobacteriaceae* from blood culture for the prevalence of extended spectrum β -lactamase (ESBL) production by the standardized disc diffusion method and confirmed by the ESBL E test strips.

Results: Ninety-five (15.8%) of the isolates were ESBL producers. Among these, 48.4% were *Klebsiella*

pneumoniae (*K. pneumoniae*) followed by 15.8% of both *Escherichia coli* (*E. coli*) and *Enterobacter cloacae* (*Ent. cloacae*). Other isolates produced ESBL in low numbers.

Conclusion: *Klebsiella pneumoniae* produced ESBL in significant numbers. Extended spectrum β -lactamase gram-negative bacilli present significant diagnostic and therapeutic challenges to the management of infections due to these organisms. Microbiology laboratories should start reporting ESBL producing *Enterobacteriaceae* organism due to their importance in respect to antibiotic therapy and infection control aspects.

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Beta-lactam antibiotics are among the safest and most frequently prescribed antimicrobial agents worldwide. The emergence of resistance to these agents in the past 2 decades has resulted in a major clinical crisis.^{1,2} Gram-negative bacteria resistant to agents such as extended-spectrum cephalosporins, monobactam, β -lactam, β -lactamase inhibitors combinations and carbapenems have emerged through the production of variety of β -lactamases, alteration in the penicillin binding proteins, outer membrane permeability, and combination of multiple mechanisms of resistance. This increase has paralleled

the introduction, administration and over use of β -lactam.³

Extended-spectrum β -lactamases (ESBLs) are primarily plasmid-mediated enzymes frequently derived from TEM or SHV-related enzyme. Both TEM-1 and SHV-1 are parent enzymes that confer resistance to ampicillin.⁴

The ESBL-producing isolates are resistant not only to amino penicillin, ureidopenicillins and narrow-spectrum cephalosporins, but all extended-spectrum cephalosporin and aztreonam.⁵ Cephamycins and carbapenem retain activity against these isolates.

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Over 120 different ESBL types have been identified, with each type conferring a slight different susceptibility profile, complicating selection of therapy.⁶

Emergence of ESBL-producing isolates has important clinical and therapeutic implication. First, in most bacterial isolates, resistance determinants for ESBL production are carried on plasmids that can be easily spread from organism to organism.⁵ Second, the spread of resistance toward extended-spectrum cephalosporins further limits the utility of the β -lactam class and may lead to increased prescription of more broad-spectrum and expensive drugs such as imipenem. In addition, these isolates may escape detection with routine susceptibility testing performed by a clinical microbiology laboratory, which can result in adverse therapeutic outcomes.^{7,8} More important, antibiotic selection for treatment of serious infection due to ESBL-producing *Enterobacteriaceae* is a clinical challenge due to the complex nature of in-vitro susceptibility testing and vivo correlation. Perhaps the biggest challenge lies in overcoming widespread unawareness among clinicians regarding these resistant organisms due to underreporting by microbiology laboratories and lack of an obvious marker to indicate production of an ESBL.⁹

Extended-spectrum β -lactamase (ESBL) producing organisms pose unique challenges to clinical microbiologists, clinicians, infection control professionals and antibacterial-discovery scientists. These ESBL-producing resistant pathogens include *Escherichia coli* (*E. coli*), *Klebsiella pneumoniae* (*K. pneumoniae*), *Enterobacter cloacae* (*Ent. cloacae*), *Serratia marcescens* (*S. marcescens*), *Citrobacter freundii* (*C. freundii*), *Pseudomonas aeruginosa* (*P. aeruginosa*) and *Acinetobacter baumannii* (*A. baumannii*).

The objective of this study was to determine the prevalence of ESBL-producing *Enterobacteriaceae* isolated from blood cultures in a tertiary care hospital.

Methods. This study was carried out at the Clinical Microbiology Department of the Armed Forces Hospital, Riyadh, Kingdom of Saudi Arabia, a 1200 bed, tertiary care facility. A total of 601 non-duplicate *Enterobacteriaceae* isolated from blood culture during the period of the study from January 2003 to December 2004 were reviewed. The isolates were identified by standard microbiological techniques, API 20E (bioMérieux, Marcy l'Etoile, France). Sensitivity testing was carried out using standardized disk diffusion method¹⁰ for ampicillin (10 μ g), amoxicillin-clavulanic acid (30 μ g), sulfamethoxazole-trimethoprim (2.5-50 μ g)

Table 1 - Frequency of *Enterobacteriaceae* producing ESBL among the isolates (N=601).

Organism	No. of isolates	Organism (%)	No. of ESBL Producers	ESBL Producers (%)
<i>Klebsiella pneumoniae</i>	206	(34.3)	46	(48.4)
<i>Escherichia coli</i>	195	(32.4)	15	(15.8)
<i>Enterobacter cloacae</i>	80	(13.3)	15	(15.8)
<i>Enterobacter aerogenes</i>	18	(3)	6	(6.3)
<i>Citrobacter freundii</i>	7	(1.2)	5	(5.3)
<i>Serratia marcescens</i>	25	(4.1)	4	(4.2)
<i>Proteus stuarti</i>	6	(1)	2	(2)
<i>Klebsiella oxytoca</i>	21	(3.5)	1	(1.1)
<i>Proteus mirabilis</i>	18	(3)	1	(1.1)
<i>Enterobacter sakazaki</i>	6	(1)	0	-
<i>Serratia liquifans</i>	4	(0.7)	0	-
<i>Salmonella species</i>	15	(2.5)	0	-
Total	601	(100)	95	(100)
ESBL - extended spectrum β -lactamase				

cefuroxime (30 μ g), ceftriaxone (30 μ g) ceftazidime (30 μ g) aztreonam (30 μ g), ciprofloxacin (5 μ g), tazobactam/piperacillin (75/10 μ g), cefepime (30 μ g), gentamicin (10 μ g), netilmicin (30 μ g), amikacin (30 μ g), imipenem (10 μ g), meropenem (10 μ g) colistin (25 μ g) (Oxoid, Basingstoke, United Kingdom) *P. aeruginosa* ATCC (27853), and *E. coli* ATCC (35218) were used as control strains.

Isolates with intermediate or resistant susceptibility for extended-spectrum cephalosporin and aztreonam were retested using ESBL E-test strip for confirmation of ESBL production (AB Biodisk, Piscataway, New Jersey). E-test strips with gradient concentration of ceftazidime (CAZ) or cefotaxime (CTX) at one end and CTX or CAZ with clavulanic acid (CA) at the other end was performed in accordance with the manufacturer recommendation. The ESBL production was confirmed by a ratio of CAZ or CTX minimum inhibitory concentration (MIC) to CAZ or CTX with CA of >8 . The ESBL production was also identified by the presence of a phantom zone, or a deformity of strip, a non-determinable (ND) was declared when MICs were greater than the range of strip.

Results. Of the 601 isolates of the members of the family *Enterobacteriaceae*, *K. pneumoniae* was the most frequent organism isolated 206 (34.3%) followed by *E. coli* 195 (32.4%), *Ent. cloacae* 80 (13.3%) and *S. marcescens* 25 (4.1%) **Table 1**. Extended spectrum β -lactamase production was confirmed by ESBL E-test strip in 95 isolates making a prevalence of 15.8%. *Klebsiella pneumoniae* was the most ESBL producer 48.4%, followed by both *E. coli* and *Ent. cloacae* each 15.8%, *Enterobacter aerogenes* (*Ent. aerogenes*) produced ESBL at lower rate at 6.3%, *C. freundii* at 5.3% and *S. marcescens* at 4.2%.

Klebsiella oxytoca, *Proteus mirabilis* (*P. mirabilis*) and *Proteus stuarti* (*P. stuarti*) produced minimal ESBL (**Table 1**).

Enterobacter sakazaki, *S. liquifans*, and *Salmonella* species did not produce any detectable ESBL. All ESBL producing isolates were sensitive to carbapenems.

Discussion. Resistance to β -lactam antimicrobial agents, especially extended-spectrum cephalosporins and other antimicrobial, among clinical isolates of gram-negative bacteria is increasing worldwide.^{11,12} Reports of clinical failure and nosocomial infections due to ESBL are emerging.¹³⁻¹⁵ Risk factors for infection with ESBL producing organisms included prolonged hospitalization, the use of various invasive diagnostic producers and prior antibiotic treatment included β -lactam agents.¹⁶

The emergence of ESBL has created not only a diagnostic problem but also poses a potential therapeutic challenge for the use of β -lactam in serious infections by the *Enterobacteriaceae*. It has also implications for nosocomial infections in intensive care unit and other special care units where the use of extended-spectrum β -lactams is high. According to a survey by the National Committee for Clinical Laboratory Standards the prevalence of ESBL is probably underestimated.¹⁷ The prevalence of ESBL-producing isolates among *Enterobacteriaceae* range from a national average of 3% in the United States¹⁸ to a much higher numbers in Europe where the prevalence of ESBL production among isolates varies greatly from country to country. In the Netherlands, a survey of 11 hospital laboratories showed that <1% of *E. coli* and *K. pneumoniae* strains possessed an ESBL.¹⁹ However, in France as many as 11.4% of *K. pneumoniae* and 47.7% of *Ent. aerogenes* were found to be ESBL producers.²⁰

The reports on ESBL production by *Enterobacteriaceae* from Saudi Arabia are few. Babay²¹ reported ESBL production in 36% of their isolates, Bilal and

Gedebou²² detected ESBL in 27.5% of *K. pneumoniae* in Abha, Kader and Kumar²³ reported ESBL in 4.8% of their gram negative isolates. The prevalence of ESBL among our isolates was 15.8%. This is lower than some reported results from Saudi Arabia.^{21,22} Higher rates were reported from India 68%²⁴ and Pakistan 45%.²⁵

We can explain the variation in the prevalence of production of ESBL among our isolates by the fact that our isolates were mainly from blood culture while in the other studies it was from different sites, wounds, urine, tracheal aspirates, and others. The majority of the ESBL producers among our isolates were *K. pneumoniae* (48.4%) and this is similar to other findings from Riyadh (42%),²¹ United States²⁶ and Argentina (48%)²⁷ where *K. pneumoniae* was the most common ESBL producers.

Escherichia coli and *Ent. cloacae* produced ESBL at similar rate 15.8% and although these are reported to be high ESBL producers by other studies,^{21,28} it did not seem to be so in our study. Other *Enterobacteriaceae* produced ESBL in low numbers.

In conclusion, data regarding the prevalence of ESBL strains in Saudi Arabia is limited.^{23,29} We can increase the level of information regarding the prevalence of ESBL by conducting studies in hospitals in Saudi Arabia, and providing data for clinicians and also for comparing the prevalence of ESBL-producing of gram-negative at both local and national levels.

References

1. Wood AJ. Antimicrobial-drug resistance. *N Eng J Med* 1996; 335: 1445-1453.
2. D'Agata FEMC. Antibiotic resistance and exposure to different generation cephalosporins. *N Eng J Med* 2000; 28: 2678-2681.
3. Fridkin SK, Steward CD, Edwards JR, Pryor ER, McGowan JE Jr, Archibald LK, et al. Surveillance of antimicrobial use and antimicrobial resistance in United States hospitals: project ICARE phase 2. Project Intensive Care Antimicrobial Resistance Epidemiology (ICARE) hospitals. *Clin Infect Dis* 1999; 29: 245-252.
4. Pitout JD, Sanders CC, Sanders WE Jr. Antimicrobial resistance with focus on β -lactam resistance in gram-negative bacilli. *Am J Med* 1997; 103: 51-59.
5. Livermore DM. β -lactamases in laboratory and clinical resistance. *Clin Microbiol Rev* 1995; 8: 557-584.
6. Jacoby GA, Bush K. Amino acid sequence for TEM, SHV and Oxa extended spectrum and inhibitor resistance β -lactamases. (Cited March 2001). Available from <http://www.lahey.org/studies/wbt.htm>.
7. Bush K. Is it important to identify extended spectrum β -lactamase-producing isolates? *Eur J Clin Microbiol Infect Dis* 1996; 15: 361-364.
8. Tenover FC, Mohammed MJ, Gorton TS, Dembek ZF. Detection and reporting of organisms producing extended spectrum β -lactamases; Survey of Laboratories in Connecticut. *J Clin Microbiol* 1999; 37: 4065-4070.

9. Paterson DL, Yu VL. Extended-Spectrum β -lactamases: a call for improved detection and control (editorial). *Clin Infect Dis* 1999; 29: 1419-1422.
10. Andrews JM. BSAC Standardized disc susceptibility testing method. *J Antimicrobial Chemother* 2000; 46 (Suppl 1): 39-52.
11. Goossens H. MYSTIC (Meropenem Yearly Susceptibility Test Information Collection) results from Europe: Comparison of antibiotic susceptibility between countries and center types. *J Antimicrob Chemother* 2000; 46 (Suppl T2): 39-52.
12. Andrews JM, BSAC Working Party On Susceptibility Testing ft. BSAC standardized susceptibility testing method. *J Antimicrob Chemother* 2001; 48 (Suppl S1): 43-57.
13. Pagani L, Perilli M, Migliavacca R, Lazzaro F, Amicosante G. Extended-Spectrum TEM- and SHV-type β -lactamase-producing *Klebsiella pneumoniae* strains causing outbreaks in intensive care units in Italy. *Eur J Clin Microbiol Infect Dis* 2000; 19: 765-772.
14. Meyer KS, Urban C, Eagan JA, Berger BJ, Rahal JJ. Nosocomial outbreak of *Klebsiella* infection resistant to late-generation cephalosporins. *Ann Intern Med* 1993; 119: 353-358.
15. Karas JA, Pillay DG, Muckart D, Sturm AW. Treatment failure due to extended spectrum β -lactamase. *J Antimicrob Chemother* 1996; 37: 203-204.
16. Quinn JP. Clinical significance of extended spectrum β -lactamases. *Eur J Clin Microbiol Infect Dis* 1994; 13 (Suppl 1): S39-42.
17. Jacoby GA, Han P. Detection of extended-spectrum β -lactamases in clinical isolates of *Klebsiella pneumoniae* and *Escherichia coli*. *J Clin Microbiol* 1996; 34: 908-911.
18. Bradford PA. Extended-spectrum β -lactamases in the 21st century: characterization, epidemiology, and detection of this important resistance threat. *Clin Microbiol Rev* 2001; 14: 933-951.
19. Stobberingh EE, Arends J, Hoogkamp-Korstanje JAA, Goossens WHF, Visser MR, Buiting AGM, et al. Occurrence of extended-spectrum β -lactamases (ESBL) in Dutch hospitals. *Infection* 1999; 27: 348-354.
20. Bell JM, Turnidge JD, Gales AC, Pfaller MA, Jones RN; Sentry APAC Study Group. Prevalence of extended-spectrum β -lactamase (ESBL)-producing clinical isolates in the Asia-Pacific region and South Africa: regional results from SENTRY Antimicrobial Surveillance Program (1998-99). *Diagn Microbiol Infect Dis* 2002; 42: 193-198.
21. Babay HA. Detection of extended β -lactamases in members of the family *Enterobacteriaceae* at a teaching hospital, Riyadh, Kingdom of Saudi Arabia. *Saudi Med J* 2002; 23: 186-190.
22. Bilal NE, Gedebo M. Clinical and community strains of *Klebsiella pneumoniae*: multiple and increasing rates of antibiotic resistance in Abha, Saudi Arabia. *Br J Biomed Sci* 2000; 57: 185-191.
23. Kader AA, Kumar KA. Prevalence of ESBL among multi-drug resistance gram-negative isolates from a general hospital in Saudi Arabia. *Saudi Med J* 2004; 25: 570-574.
24. Mathur P, Kapil A, Das B, Dhawan B. Prevalence of extended spectrum β -lactamase producing Gram negative bacteria in tertiary care hospital. *Indian J Med Res* 2002; 115: 153-157.
25. Shah AA, Hasan F, Ahmed S, Hameed A. Extended-spectrum β -lactamases in *Enterobacteriaceae*: related to age and gender. *New Microbiol* 2002; 25: 363-366.
26. Coudron PE, Moland ES, Sanders CC. Occurrence and detection of Extended-Spectrum β -Lactamase in members of *Enterobacteriaceae* of a veterans Medical Center: Seek and You May Find. *J Clin Microbiol* 1997; 35: 2593-2597.
27. Bantar C, Famiglietti A, Goldberg M. Three-year surveillance study of nosocomial bacterial resistance in Argentina. The Antimicrobial committee; and National Surveillance Program (SIR) Participants Group. *Int J Infect Dis* 2000; 4: 85-90.
28. Albertini MT, Benoit C, Berardi L, Berrouane Y, Boisivon A, Cahen P, et al. Microbiology Surveillance Network of Northern France. Surveillance of methicillin-resistance *Staphylococcus aureus* (MRSA) and *Enterobacteriaceae* producing extended-spectrum β -lactamase (ESBL) in Northern France: a five-year multicentre incidence study. *J Hosp Infect* 2002; 52: 107-113.
29. Ramadan MA, Tawfik AF, Shibl AM. Effect of β -lactamase expression on susceptibility of local isolates of *Enterobacter cloacae*, *Serratia marcescens* and *Pseudomonas aeruginosa* to β -lactam antibiotics. *Chemotherapy* 1995; 41:193-199.