# Manipulation of flaxseed inhibits tumor necrosis factoralpha and interleukin-6 production in ovarian-induced osteoporosis

Hala M. Abdelkarem, PhD, Madeha M. Abd El-Kader, PhD, Seham A. Kasem, PhD,

## ABSTRACT

الأهداف: تقييم تأثير الحبوب الكاملة من بذور الكتان، وزيت بذور الكتان على السيتوكينات المولدة للالتهاب وذلك في الفئران التي أُجريت لها عملية استئصال المبيض، وكذلك تأثير مثل هذه الأغذية على مرض هشاشة العظام.

الطريقة : أجريت هذه الدراسة في قسم الغذاء والتغذية بكلية الزراعة، جامعة الملك سعود، الرياض، المملكة العربية السعودية وذلك خلال الفترة من أكتوبر إلى ديسمبر 2009م. شملت الدراسة 48 جرذياً من النوع دوللي سبراغو والتي تبلغ أعمارها 3 أشهر، وقد تم تقسيمها عشوائياً إلى المجموعات التالية : المجموعة 1 وهي التي أُطعمت طعاماً خادعاً مع تزويدها بحمية محددة، والمجموعة 2 وأجريت لها عملية استئصال المبايض مع حمية أساسية، والمجموعة 3 وأجريت لها نفس العملية مع إعطاؤها 200 من بذور الكتان، والمجموعة 4 وأجريت لها العملية مع 400 من بذور الكتان، والمجموعة 5 أجريت لها نفس العملية مع 40% من بذور الكتان، والمجموعة 5 أجريت لها نفس العملية ولكن مع 50 من زيت بذور الكتان، والمجموعة موتع ليا العملية مع 40% من بذور الكتان، والمجموعة 5 أجريت الما نفس العملية مع 10% من زيت بذور الكتان، والمجموعة 4 موتع ليا العملية مع 40% من بذور الكتان، والمجموعة 5 موتع ليا العملية مع 40% من بذور الكتان، والمجموعة 4 الما نفس العملية مع 50% من زيت بذور الكتان، والمجموعة 5 موتع ليا العملية مع 10% من زيت بذور الكتان، والمجموعة 5 موتع ليا العملية مع 10% من زيت بذور الكتان، والمجموعة 5 موتع ليا العملية مع 10% من زيت بذور الكتان، والمجموعة 5 موت ليا العملية مع 10% من زيت بذور الكتان، والمجموعة 10 معدلات كلاً من: ألكالين فوسفاتيز، وانترليوكين 6، وعامل تنخر الأورام ألفا، والكالسيوم، والفسفور، والمعنيسيوم.

النتائج: أشارت نتائج الدراسة إلى زيادة معدلات انترليوكين-6، وعامل تنخر الأورام ألفا في الجرذان التي خضعت لعملية استئصال المبايض مقارنةً بالمجموعة 1، في حين لم يكن هناك اختلافاً ظاهراً في معدلات كلاً من: ألكالين فوسفاتيز، والكالسيوم، والفسفور، والمغنيسيوم بين كافة المجموعات المشاركة في التجربة. وكان هناك نقصاً واضحاً في معدلات انترليوكين-6، وعامل تنخر الأورام ألفا وذلك في المجموعات التي احتوت حميتها على بذور الكتان (المجموعة 3 و4) وكذلك المجموعات التي تناولت زيت بذور الكتان (المجموعة 5 و6).

**خاتمة**: اقترحت الدراسة إمكانية تأثير بذور وزيت الكتان على الوقاية من مرض هشاشة العظام الناتج عن نقص هرمون الاستروجين وذلك عن طريق خفض عمليات هدم العظام. ونحن بحاجة إلى المزيد من الدراسات من أجل تقييم عمليات الأيض لمثل هذه الأغذية وكيفية تأثيرها على عمليات هدم وبناء العظام.

**Objectives:** To evaluate the potential effects of whole flaxseed (FS), and/or flax oil (FO) incorporation into the diet on the level of pro-inflammatory cytokines in ovariectomized (OVX) rats model of osteoporosis.

Methods: This study was performed in the Food Science & Agriculture Collage, King Saud University, Kingdom of Saudi Arabia from October to December 2009. Fortyeight, 3-month-old female Sprague-Dawley rats were randomly divided into 6 groups: Group 1 - sham + control diet; Group 2 - OVX rats + basal diet; Group 3 - OVX + 20% whole FS; Group 4 - OVX rats + 40% FS; Group 5 - OVX rats + 5% FO; Group 6 - OVX rats + 10% FO. All OVX rats underwent bilateral ovariectomy. The experiment was continued for 2 months. Serum bone alkaline phosphatase (B-ALP), interleukin-6 (IL-6), tumor necrosis factor-alpha (TNF-α), calcium (Ca), phosphorous (P), and magnesium (Mg) were measured.

**Results:** A significant increase of serum IL-6 and TNF- $\alpha$  concentrations were observed between OVX rats when compared with Group 1, while there was no significant difference in the activity of B-ALP, serum Ca, P, and Mg among all groups. A remarkable significant decrease of serum levels of IL-6 and TNF- $\alpha$  was observed in the group of rats that were fed with FS (Groups 3 and 4) and FO (Groups 5 and 6).

**Conclusion:** This study suggests that FS and FO might be useful in the prevention of estrogen-deficiency induced osteoporosis via decreasing osteoclastogenesis. Further studies are needed to demonstrate their efficacy in humans by using bioactive components of FS, and to clarify their mechanism of action.

#### Saudi Med J 2011; Vol. 32 (4): 369-375

From the Food Science and Nutrition Department (Abdelkarem), Food Science & Agriculture Collage, King Saud University, Kingdom of Saudi Arabia, and the Nutrition Department (Abd El-Kader, Kasem), National Research Center, Cairo, Egypt.

Received 4th October 2010. Accepted 14th February 2011.

Address correspondence and reprint request to: Prof. Dr. Hala M. Abdelkarem, Biochemical Nutrition, Food Science and Nutrition Department, Food Science & Agriculture Collage, King Saud University, PO Box 22452, Riyadh 11495, Kingdom of Saudi Arabia. Tel. +966 (1) 530707977. Fax: +966 (1) 4775406. E-mail: halaabdelkarem@yahoo.com

varian hormone deficiency is a major risk factor Jfor osteoporosis in postmenopausal women.<sup>1,2</sup> Estrogen replacement therapy (ERT) has long been used to alleviate postmenopausal symptoms and lowers the risk of osteoporosis. However, estrogen treatment is associated with a higher risk of certain types of cancer or contraindications.<sup>3,4</sup> As estrogen inhibits bone resorption, its deficiency will increase bone loss.<sup>5</sup> Estrogen down-regulates bone-resorbing cytokines, such as tumor necrosis factor-alpha (TNF- $\alpha$ ), interleukin (IL)-6, and prostaglandin (PG) E2.6 Phytoestrogens, are compounds found in plants and plant products that possess some estrogenic or antiestrogenic activity. Recent evidence suggests that phytoestrogens are non-steroidal plant compounds found in many fruits, vegetables, and grains. These phytoestrogens may act as estrogen antagonists in mammary gland, and in contrast, they act as estrogen agonists in bone.<sup>7</sup> Lignans, one type of phytoestrogen, are diphenolic compounds similar in structure to endogenous non-steroid hormones, and are hypothesized to act in vivo to alter hormone metabolism and subsequent bone metabolism.<sup>8</sup> Flaxseed (FS), one of the edible plant foods is the highest rich source of lignans, which are reported to have both weak estrogenic and anti-estrogenic activities.9 Flaxseed is also a rich source of polyunsaturated fatty acid (PUFA), especially linolenic acid (18:3 [n-3]).<sup>10</sup> Alpha linolenic acid (ALA) may decrease the rate of bone resorption by inhibiting the biosynthesis of cytokines, such as PG, ILs and TNF.11 Lignans present in FS may also possess antioxidant properties. Oxygen-derived free radicals, resulting from excessive production of reactive oxygen species perturb the normal redox balance of osteogenesis including bone formation and resorption. These findings indicate that free radicals have marked capacity to degenerate the bone metabolism, and enhance osteoclast formation and bone resorption. A previous study<sup>12</sup> demonstrated that oxidative stress is involved in the pathogenesis of bone loss in female rats due to chronic inflammatory diseases, aging, and osteoporosis. Therefore, FS may reduce the rapid rate of bone loss experienced by postmenopausal women in part, by enhancing antioxidant status and exerting a positive effect on the bone.<sup>13</sup> In a follow up study,<sup>13</sup> FS can potentially exert positive effects on bone of postmenopausal women. They assigned 60 postmenopausal women not on hormone replacement therapy receiving either 40 g FS, or 40 g wheat as control supplement for 3 months, and showed no amelioration

**Disclosure**. The authors have no conflicting interests, not supported, or funded by any drug company

of serum and urinary biomarkers of bone metabolism in the early evaluation, or whether longer-term study using bioactive components of FS, such as lignans or its oil can elucidate a positive influence on BMD. The n-3 PUFAs have anti-inflammatory properties that was mediated by the production of anti-inflammatory eicosanoids, which in turn offset the production of pro-inflammatory eicosanoids through competitive inhibition within their common metabolic pathways.<sup>14</sup> Limiting the production of pro-inflammatory eicosanoids, such as PGE2 may be an important factor in minimizing the production of IL-6, IL-1, and TNF- $\alpha$ , which may mediate inflammation-associated bone abnormalities.<sup>15</sup> The FS derived ALA has been shown to decrease in vivo PGE2 concentrations in rat bones.<sup>16</sup> In addition, flaxseed oil (FO) has been shown to decrease TNF- $\alpha$ and IL-1 in human peripheral blood mononuclear cells. Modulation of the dietary n-6 to n-3 ratio has been shown to be beneficial in various clinical inflammatory diseases,<sup>17,18</sup> and animal models of bone metabolism.<sup>19-</sup> <sup>21</sup> Little published data indicated a response of bone metabolism during intestinal inflammation to a diet rich in ALAs derived from FO. This information initiated our interest to examine the effect of dietary intake of FS and/or FO as therapeutic functional foods on proinflammation cytokines induced in ovariectomized (OVX) rats model osteoporosis.

**Methods.** Animals. This study was performed in the Food Science & Agriculture Collage, King Saud University, Kingdom of Saudi Arabia from October to December 2009. Forty-eight female Sprague-Dawley rats aged 3 months, and weighing between 180-250 g were obtained from the university animal facility. Throughout the experiment, rats were housed in stainless steel cages with available water ad libitum. All animals were kept under normal healthy conditions and fed basal diet.

Experiment procedure. Rats were randomly divided into 6 groups (n=8 each). After one week of habituation to the facilities, the animals were used for the study. All OVX rats underwent bilateral ovariectomy via a dorsal approach with a small midline dorsal skin incision, the sham-operation rats (Group 1) were subjected to sham surgery exposure without removing the ovaries. After 2 weeks of recovery from the operation, Group 1 and OVX control group (Group 2) received basal diet without any addition of FS or FO. The other 4 groups were: Group 3 - OVX + 20% FS diet; Group 4 - OVX + 40% FS diet; Group 5 - OVX + 5% FO diet; and Group 6 - OVX + FO 10% (Table 1). There were 8 rats in each group. Dietary treatment was started 2 weeks post-ovariectomy, and continued for 2 months. After 2 months, the animals were sacrificed by cervical

dislocation under anesthesia; blood samples were obtained, and left to clot. Serum was separated after centrifugation at 3500 rpm for 15 minutes, and frozen at -70 °C until analysis.

Analytic methods. Serum B-ALP (a marker of bone formation), IL-6 and TNF- $\alpha$  (marker of bone resorption) were measured using enzyme linked immunosorbent assay (ELISA) method kits, B-ALP [AviBion, Orgenium Laboratories Division, Vantaa, Finland], TNF- $\alpha$  & IL-6 ELISA [Ani Biotech Oy, Orgenium Laboratories Division, Vantaa, Finland). The reading was carried out using ELISA microplate reader (VERSA Max, Molecular Devices Corporation, MN, USA). Serum calcium (Ca), phosphorus (P), and magnesium (Mg) were measured by enzymatic method as cited in the United Diagnostics Industry.<sup>22,23</sup> This study was approved by the IBR at the King Saud University of Kingdom Saudi Arabia.

Statistical analyses were carried out using 2-way analysis of variance (ANOVA), and paired t-test for

normally-distributed samples. Data were analyzed using the Statistical Package for Social Sciences version 17.1 (SPSS Inc, Chicago, IL, USA). *P*<0.05 was considered significant.

**Results.** Effects of ovariectomy, FS and FO on food intake, body weight, and relative organ weight. Data on food intake, body weights and organ weights are shown in Table 2. The average food intake in Group 2 was significantly higher from Group 1 and other treated groups. The weight gains of rats in Group 2 were significantly higher (38%) than those of Group 1. The OVX rats showed atrophy of uterine tissues, however, there were no differences in the liver weight among the study groups (Table 2).

Effects of ovariectomy, FS, and FO on biochemical markers. The OVX, a known stimulus of bone resorption, induced a significant increase in each markers of pro-inflammation cytokines (IL-6 & TNF- $\alpha$ ), above the

**Table 1** - Composition of the control diet based on American Institute of Nutrition (AIN 93)<sup>22</sup> standard diet.

		Flax				
Components	Control	Flax powder	Flax powder	Flax oil	Flax oil	
Casein	200	200	200	200	200	
Cornstarch	367.50	167.5	0	367.50	367.48	
Dyetrose (Dextrinized cornstarch (90-94% tetrasaccharides)	132	132	132	132	132	
Sucrose	100	100	100	100	100	
Cellulose	50	50	15	50	50	
Corn oil	100	100	100	50	0	
Flaxseed powder	0	200	400	0	0	
Flaxseed oil	0	0	0	50	100	
Butylhydroquinone	0	0.02	0.02	0.02	0.02	
l-Cystine	3.0	3.0	3.0	3.0	3.0	
Choline bitartrate	2.5	2.5	2.5	2.5	2.5	
Mineral mix	35	35	35	35	35	
Vitamin mix	10	10	10	10	10	
Both diets were modified A	IN93G standard ro	dent diet and co	ntained either 2	20, 40% FS or	5, 10% FS	

Table 2 - Effects on ovariectomized rats whole flaxseed and flax oil on food intake, body weight, and relative organ weight.

D .	Group 1	Group 2	Group 3	Group 4	Group 5	Group 6
Parameters			Flaxseed		Flax oil	
<i>Food intake, g/day/rat</i> <i>P</i> -value	10.84 ± 0.41	15.38 ± 0.40 0.001	13.16 ± 0.21 0.008	11.07 ± 0.49 0.003	12.05 ± 0.63 0.01	11.04 ± 0.82 0.004
Body weight, g						
Initial	155.4 ± 5.03	154.6 ± 9.04	154.0 ± 8.14	154.2 ± 6.73	154 ± 8.97	154.2 ± 8.54
Final	210 ± 6.98	242.6 ± 10.93	245.4 ± 8.33	265.2 ± 9.69	235.6 ± 7.82	234.4 ± 10.39
P-value		0.0001	0.0001	0.0001	0.0001	0.0001
Organ weight*						
Uterus	$0.32 \pm 0.03$	$0.21 \pm 0.05$	$0.19 \pm 0.03$	$0.18 \pm 0.04$	$0.20 \pm 0.04$	$0.19 \pm 0.06$
Liver	5.22 ± 0.356	4.90 ± 0.293	$5.40 \pm 0.174$	5.86 ± 0.122	4.74 ± 0.29	$4.60 \pm 0.354$

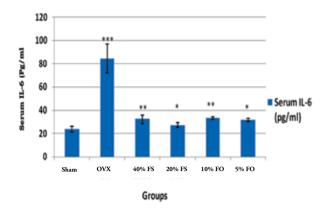
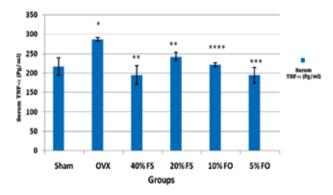


Figure 1 - Effect of dietary whole flaxseed (FS) and flax oil (FO) on serum interleukin (IL)-6. The IL-6 was determined by enzyme-linked immunosorbent assay method, ovariectomized rats (OVX) group compared with the Sham group. After treatment, FS groups (20% and 40%) compared with OVX group and FO groups (5% and 10%) compared with OVX group for 2 months. The plotted data represent the mean ± standard error for each dietary treatment group. For statistical differences between treatments \*p=0.03, \*\*p=0.02, \*\*\*p= 0.001



**Figure 2** - Effect of dietary whole flaxseed (FS) and flax oil (FO) on serum tumor necrosis factor-alpha (TNF-α). The TNF-α was determined by enzyme-linked immunosorbent assay method. The ovariectomized rats (OVX) group was compared with the Sham group. After treatment, the FS groups (20% and 40%) was compared with the OVX groups, and FO groups (5% and 10%) were compared with the OVX group for 2 months. The plotted data represent the mean ± standard error for each dietary treatment group. For statistical differences between treatments. \**p*=0.04, \*\**p*=0.03, \*\*\**p*=0.02, \*\*\*\**p*=0.0001

levels measured in Group 1. Thus, the mean value of serum IL-6 was 84.40 ±12.18 pg/ml in Group 2 versus  $23.78 \pm 2.62 \text{ pg/ml}$  in Group 1 (p=0.01) (Figure 1), and that of TNF- $\alpha$  was 287.62 ±3.80 pg/ml in Group 2 versus 217.48  $\pm$  22.19 pg/ml in Group 1 (p=0.04) (Figure 2). No significant difference was observed in the activity of serum B-ALP of Group 2 when compared with Group 1 (Table 3). For each marker, whole FS at Group 3, and Group 4 reversed the effect of OVX (Figures 1 & 2). Thus, the mean values of serum IL-6 was 32.33 ± 3.70 pg/ml (p=0.01) for Group 3, and  $27.23 \pm 2.03 \text{ pg/ml} (p=0.02)$  for Group 4, and that of serum TNF- $\alpha$  was 195.50 ± 24.37 pg/ml (*p*=0.03) for Group 3, and 242.86  $\pm$  0.78 pg/ml (*p*=0.03) for Group 4. The treatment of OVX rats significantly decreased the level of serum IL-6 in Group 5  $(33.33 \pm 1.19 \text{ pg/ml})$ ; p=0.03), and in Group 6 (31.60 ± 1.30 pg/ml; p=0.02), and serum TNF- $\alpha$  in Group 5 was 221.90 ± 4.70 pg/ml (p=0.03), and 194.96 ± 20.43 pg/ml (p=0.02) in Group 6 (Figures 1 & 2). There was no significant difference in the activity of serum B-ALP, Ca, P, and Mg among the different groups (Table 3).

**Discussion.** The present study showed positive effects of dietary FS on biochemical markers of bone remodeling in OVX rats model of osteoporosis.24 It showed that the body weight significantly increased (63.1%), while the uterine weight decreased greatly (34.4 %) of OVX rats when compared with Group 1 (Table 2). This indicates that the animals had became estrogen deficient. The administration of FS rich in phytoestrogen at different concentrations has inhibited the increase of body weight, and did not affect the uterine weight.<sup>24</sup> The present data are consistent with some reports,<sup>26,27</sup> that has attributed the mechanism of action of some phytoestrogen to their high binding affinities to the intercellular estrogen receptors. They supported that these compounds may act in target tissues as agonist or antagonists in the absence of endogenous estradiol. This is in agreement with our results indicating that the weight of the uterus of the treated group did not differ significantly from that of the untreated groups.

Table 3 - The levels of biochemical parameters in ovariectomized rats, fed with whole flaxseed and flax oil after 2 months treatment.

11.15 ± 0.28 5.13 + 0.26	Flax 10.54 ± 0.33 45.17 ± 0.14	11.23 ± 0.21	11.08 ± 0.20	<b>x oil</b> 11.15 ± 0.34
$5.13 \pm 0.26$	$45.17 \pm 0.14$	(72 0 17	101 011	
J115 ± 0120	$\pm 0.17 \pm 0.14$	$4.73 \pm 0.17$	4.94 ± 0.16	$4.75 \pm 0.12$
$2.61 \pm 0.02$	$2.62 \pm 0.07$	$2.65 \pm 0.14$	$2.65\pm0.04$	$2.61 \pm 0.04$
$20.30 \pm 0.51$	$21.78 \pm 1.84$	$22.48 \pm 1.30$	$18.94 \pm 0.85$	19.08 ± 1.31
	20.30 ± 0.51	$20.30 \pm 0.51 \qquad 21.78 \pm 1.84$	20.30 ± 0.51 21.78 ± 1.84 22.48 ± 1.30	

The obtained data showed that the OVX rats have significantly increased in biochemical parameters as observed with the increase of pro-inflammatory activity.<sup>23</sup> These results could be supported by the fact that pro-inflammatory cytokines are responsible for osteoclastogenesis and increased trabecular bone resorption after loss of sex steroids due to menopause.<sup>28,29</sup> The concentrations of serum Ca, P, and Mg showed no significant difference after ovariectomy, and these results are inconsistent with previous study<sup>30</sup> that serum Ca decreased in the OVX rats. In this study, 2 months treatment of OVX with whole FS and FO revealed significant decrease of serum IL-6 and TNF- $\alpha$  (markers of bone resorption) and no significant difference of serum B-ALP activity (a marker of bone formation), therefore a reduction of osteoclastic activity and establishment of osteoblastic activity, indicate down regulation of the bone turnover rate in OVX rats.

Flaxseeds is a rich source of lignans with diphenolic ring structures resembling those of endogenous estrogens that have potential weak estrogenic and antiestrogenic activity similar to that of the isoflavones found in soy.<sup>31,32</sup> Therefore, lignans as well as other phytoestrogens including daidzein and genistein,<sup>33</sup> were proven to have an anabolic effect on bone metabolism and prevented bone loss. The TNF- $\alpha$  stimulates osteoblasts to secrete other cytokines (IL-1B, IL-6) and PGE2, as well as osteoclasts to cause bone resorption.34,33 Estrogen inhibits the IL-1, and TNF- $\alpha$  stimulated biosynthesis of IL-6 in stromal and osteoblastic cells.36,37 It is hypothesized that suppression of the stimulatory effect of estrogen deficiency on bone metabolism is through the blocking any one of these cytokines.<sup>38</sup> This could help us to suggest that mechanism of action is possibly that the lignans in FS affect the bone through estrogen receptors.39

Several studies also suggested that FS lignans and soy isoflavones interfere with the normal physiologic activity and metabolism of estrogens, together with the ability to modulate estrogen metabolism that affect tissue exposure to biologically active estrogens (estradiol and 16-OHE1), which may inhibit pro-inflammation (IL-6 and TNF- $\alpha$ ) and osteoclastogenesis.<sup>40,41</sup> There was an association between receptor activator of nuclear factor kappa-B ligand (RANKL) and the direct or indirect effect of TNF- $\alpha$  on osteoclastogenesis. The RANKL stimulate TNF- $\alpha$  expression in osteoclast precursor cells and homogenous populations of Raw264.7 cells.<sup>42</sup> The antioxidant and free radical scavenging properties of polyphenolic compounds in several plants has recently been reported.<sup>43,44</sup> Redox oxygen scavenges (ROS) are known to induce TNF- $\alpha$  expression, suggesting that ROS acts by increasing intracellular signals that induce TNF- $\alpha$  expression, rather than by augmenting signals that stimulate osteoclast formation and function.<sup>45,46</sup> Thus, in the present results, it can be suggested that TNF- $\alpha$  production in response to RANKL was reduced by FS lignan as polyphenolic compounds, and hence, FS were involved in affecting the viability and proliferation of osteoclasts, which may be associated with regulation of bone remodeling.

Flaxseed is also a very rich source of ALA,<sup>47</sup> which is known to decrease bone turnover and increase bone mineral density in the femur and lumber bones.<sup>48</sup> The FO contains approximately 56% ALA,<sup>49</sup> a precursor of eicosapentaenoic acid (EPA), has generated interest as a potential anti-inflammatory agent due to the ability of the ALA to be converted to EPA in human and animals.<sup>30</sup> The EPA-enriched diet is potent in preventing estrogen deficiency bone loss in OVX rats.<sup>31</sup> Thus, the present study suggested that FO might contribute to the prevention of osteoporosis by the beneficial effect on cytokines and prostaglandins production, and in consequent bone resorption markers. The limitation of the study: cannot be applied on human, difficult in volunteers and consents from the target people.

In conclusion, this study is encouraging and may be promising for the consumption of FS lignans as a potential alternative therapy to prevent osteoporosis associated ovarian deficient-women. However, the biological effects observed can attribute to particular contents, as many compounds are present in FS. Further studies are needed to demonstrate their efficacy in humans by using bioactive components of FS, and to clarify their mechanism of action.

**Acknowledgment.** The authors gratefully acknowledge the Deanship of Academic-Women Students, Medical Studies and Science Sections, Research Center, King Saud University in helping us for the funding and publication.

## References

- 1. Pietschmann P, Rauner M, Sipos W, Kerschan-Schindl K. Osteoporosis: an age-related and gender-specific disease--a mini-review. *Gerontology* 2009; 55: 3-12.
- Tremollieres F, Ribot C. Bone mineral density and prediction of non-osteoporotic disease. *Maturitas* 2010; 65: 348-351.
- 3. Clarkson TB. Estrogen effects on arteries vary with stage of reproductive life and extent of subclinical atherosclerosis progression. *Menopause* 2007; 14: 373-384.
- Stanczyk FZ, Lee JS, Santen RJ. Standardization of steroid hormone assays: why, how, and when?. *Cancer Epidemiol Biomarkers Prev* 2007; 16: 1713-1729.
- Zaidi M, Blair HC, Moonga BS, Abe E, Huang CL. Osteoclastogenesis, bone resorption, and osteoclast-based therapeutics. *J Bone Miner Res* 2003; 18: 599-609.
- Weitzmann MN, Roggia C, Toraldo G, Weitzmann L, Pacifi R. Increased production of interleukin-7 uncouples bone formation from bone resorption during oestrogen deficiency. J Clin Invest 2002; 110: 1643-1650.

- Effenberger KE, Johnsen SA, Monroe DG, Spelsberg TC, Westendorf JJ. Regulation of osteoblastic phenotype and gene expression by hop-derived phytoestrogens. *J Steroid Biochem Mol Biol* 2005; 96: 387-399.
- Hutchins AM, Martini MC, Olson BA, Thomas W, Slavin JL. Flaxseed influences urinary lignan excretion in a dose-dependent manner in postmenopausal women. *Cancer Epidemiol Biomarkers Prev* 2000; 9: 1113-1118.
- Chen J, Saggar JK, Corey P, Thompson LU. Flaxseed and pure ecoisolariciresinol diglucoside, but not flaxseed hull, reduce human breast tumor growth (MCF-7) in athymic mice. *J Nutr* 2009; 139: 2061-2066.
- Ander BP, Weber AR, Rampersad PP, Gilchrist JS, Pierce GN, Lukas A. Dietary flaxseed protects against ventricular fibrillation induced by ischemia-reperfusion in normal and hypercholesterolemic rabbits. *J Nutr* 2004; 34: 3250-3256.
- 11. Griel AE, Kris-Etherton PM, Hilpert KF, Zhao G, West SG, Corwin RL. An increase in dietary n-3 fatty acids decreases a marker of bone resorption in humans. *Nutr J* 2007; 6: 2.
- Bahram H, Arjmandi, RD. The Role of Phytoestrogens in the Prevention and Treatment of Osteoporosis in Ovarian Hormone Deficiency. *Journal of the American College of Nutrition* 2001; 20: S398-S402.
- Brooks JD, Ward WE, Lewis JE, Hilditch J, Nickell L, Wong E, et al. Supplementation with flaxseed alters estrogen metabolism in postmenopausal women to a greater extent than does supplementation with an equal amount of soy. *Am J Clin Nutr* 2004; 79: 318-325.
- Wallace FA, Miles EA, Calder PC. Comparison of the effects of linseed oil and different doses of fish oil on mononuclear cell function in healthy human subjects. *Br J Nutr* 2003; 89: 679-689.
- Watkins BA, Li Y, Seifert MF. Nutraceutical fatty acids as biochemical and molecular modulators of skeletal biology. *Journal of the American College of Nutrition* 2001; 20: S410-S416.
- Weiler H, Kovacs H, Nitschmann E, Fitzpatrick Wong S, Bankovic-Calic N, Ogborn M. Elevated bone turnover in rat polycystic kidney disease is not due to prostaglandin E2. *Pediatr Nephrol* 2002; 17: 795-799.
- Liu Y, Gong L, Li D, Feng Z, Zhao L, Dong T. Effects of fish oil on lymphocyte proliferation, cytokine production and intracellular signaling in weanling pigs. *Arch Tierernahr* 2003; 57: 151-165.
- Nauroth JM, Liu YC, Van Elswyk M, Bell R, Hall EB, Chung G, et al. Docosahexaenoic acid (DHA) and docosapentaenoic acid (DPAn-6) algal oils reduce inflammatory mediators in human peripheral mononuclear cells in vitro and paw edema in vivo. *Lipids* 2010; 45: 375-384.
- Vieira de Barros K, Gomes de Abreu G, Navarro Xavier RA, Real Martinez CA, Ribeiro ML, Gambero A, et al. Effects of a high fat or a balanced omega 3/omega 6 diet on cytokines levels and DNA damage in experimental colitis. *Nutr* 2011; 27; 221-226.
- Sun D, Krishnan A, Zaman K, Lawrence R, Bhattacharya A, Fernandes G. Dietary n-3 fatty acids decrease osteoclastogenesis and loss of bone mass in ovariectomized mice. *J Bone Miner Res* 2003; 18: 1206-1216.
- Shen CL, Yeh JK, Samathanam C, Cao JJ, Stoecker BJ, Dagda RY, et al. Green tea polyphenols attenuate deterioration of bone microarchitecture in female rats with systemic chronic inflammation. *Osteoporos Int* 2011; 22: 327-337.
- 22. Gimblet EG, Marney AF, Bonsnes RW. Determination of calcium and magnesium in serum, urine, diet and stool by atomic absorption spectrophotometry. *Clin Chem* 1967; 13: 204-214.

- Gomori G. Modification of colorimetric phosphorus determination of use with photoelectric colorimeter. *J Lab Clin Med* 1942; 17: 955-960.
- Rahman MM, Bhattacharya A, Fernandes G. Docosahexaenoic acid is more potent inhibitor of osteoclast differentiation in RAW 264.7 cells than eicosapentaenoic acid. *J Cell Physiol* 2008; 214: 201-209.
- 25. Boulbaroud S, Arfaoui A, Abdelkrim C, Mesfioui A, Ouichou A, El Hessni A. Does flaxseed uptake reverse induced-loss in ovariectomized rats. *International Journal of Osteoporosis and Metabolic Disorders* 2008; 1: 24-30.
- Griffith JF, Wang YX, Zhou H, Kwong WH, Wong WT, Sun YL, et al. Reduced bone perfusion in osteoporosis: likely causes in an ovariectomy rat model. *Radiology* 2010; 254: 739-746.
- 27. Prasain JK, Arabshahi A, Moore DR 2nd, Greendale GA, Wyss JM, Barnes S. Simultaneous determination of 11 phytoestrogens in human serum using a 2 min liquid chromatography/tandem mass spectrometry method. J Chromatogr B Analyt Technol Biomed Life Sci 2010; 878: 994-1002.
- Mayer Y, Balbir-Gurman A, Machtei EE. Anti-tumor necrosis factor-alpha therapy and periodontal parameters in patients with rheumatoid arthritis. *J Periodontol* 2009; 80: 1414-1420.
- 29. Ershler WB, Keller ET. Age-associated increased interleukin-6 gene expression, late-life diseases, and frailty. *Annu Rev Med* 2000; 51: 245-270.
- Masayoshi Y. Regulatory mechanism of food factors in bone metabolism and prevention of osteoporosis. *Yakugaku Zasshi* 2006; 126: 1117-1137.
- Passeri G, Vescovini R, Sansoni P, Galli C, Franceschi C, Passeri M, et al. Calcium metabolism and vitamin D in the extreme longevity. *Exp Gerontol* 2008; 43: 79-87.
- 32. Dixon RA. Phytoestrogens. Annu Rev Plant Biol 2004; 55: 225-261.
- Sugimoto E, Yamaguchi M. Anabolic effect of genistein in osteoblastic MC3T3-E1 cells. *Int J Mol Med* 2000; 5: 515-520.
- 34. Nagata A, Tanaka T, Minezawa A, Poyurovsky M, Mayama T, Suzuki S, et al. cAMP activation by PACAP/VIP stimulates IL-6 release and inhibits osteoblastic differentiation through VPAC2 receptor in osteoblastic MC3T3 cells. *J Cell Physiol* 2009; 221: 75-83.
- 35. McLean RR. Proinflammatory cytokines and osteoporosis. *Curr Osteoporos Rep* 2009; 7: 134-139.
- 36. Gatson JW, Maass DL, Simpkins JW, Idris AH, Minei JP, Wigginton JG. Estrogen treatment following severe burn injury reduces brain inflammation and apoptotic signaling. J *Neuroinflammation* 2009; 6: 30.
- 37. Gomez CR, Plackett TP, Kovacs EJ. Aging and estrogen: modulation of inflammatory responses after injury. *Exp Gerontol* 2007; 42: 451-456.
- Lindahl G, Saarinen N, Abrahamsson A, Dabrosin C. Tamoxifen, flaxseed, and the lignan enterolactone increase stroma- and cancer cell-derived IL-1Ra and decrease tumor angiogenesis in estrogen-dependent breast cancer. *Cancer Res* 2011; 71: 51-60.
- Le Bail JC, Champavier Y, Chulia AJ, Habrioux G. Effects of phytoestrogens on aromatase, 3beta and 17beta-hydroxysteroid dehydrogenase activities and human breast cancer cells. *Life Sci* 2000; 66: 1281-1291.
- Karkola S, Wähälä K. The binding of lignans, flavonoids and coumestrol to CYP450 aromatase: a molecular modeling study. *Mol Cell Endocrinol* 2009; 301: 235-244.
- Wang LQ. Mammalian phytoestrogens: enterodiol and enterolactone. J Chromatogr B Analyt Technol Biomed Life Sci 2002; 777: 289-309.

- 42. Stolina M, Schett G, Dwyer D, Vonderfecht S, Middleton S, Duryea D, et al. RANKL inhibition by osteoprotegerin prevents bone loss without affecting local or systemic inflammation parameters in two rat arthritis models: comparison with anti-TNF alpha or anti-IL-1 therapies. *Arthritis Res Ther* 2009; 11: R187.
- 43. Nakao A, Fukushima H, Kajiya H, Ozeki S, Okabe K. RANKL stimulated TNF-alpha production in osteoclast precursor cells promotes osteoclastogenesis by modulating RANK signaling pathways. *Biochem Biophys Res Commun* 2007; 357: 945-950.
- 44. Niculescu MD, Pop EA, Fischer LM, Zeisel SH. Dietary isoflavones differentially induce gene expression changes in lymphocytes from postmenopausal women who form equol as compared with those who do not. *J Nutr Biochem* 2007; 18: 380-390.
- Ishimi Y. Soybean isoflavones in bone health. Forum Nutr 2009; 61: 104-116.
- Jagger CJ, Lean JM, Davies JT, Chambers TJ. Tumor necrosis factor-alpha mediates osteopenia caused by depletion of antioxidants. *Endocrinol* 2005; 146: 113-118.

- Kyung TW, Lee JE, Shin HH, Choi HS. Rutin inhibits osteoclast formation by decreasing reactive oxygen species and TNF-alpha by inhibiting activation of NF-kappaB. *Exp Mol Med* 2008; 40: 52-58.
- Tarpila S, Aro A, Salminen I, Tarpila A, Kleemola P, Akkila J, et al. The effect of flaxseed supplementation in processed foods on serum fatty acids and enterolactone. *Eur J Clin Nutr* 2002; 56: 157-165.
- Griel AE, Kris-Etherton PM, Hilpert KF, Zhao G, West SG, Corwin RL. An increase in dietary n-3 fatty acids decreases a marker of bone resorption in humans. *Nutr J* 2007; 6: 2.
- 50. Li Y, Greiner RS, Salem N, Watins BA. Impact of dietary n-3 fatty acid deficiency on rat bone tissue fatty acid composition. *Lipids* 2003; 38: 683-686.
- 51. Farmer C, Giguère A, Lessard M. Dietary supplementation with different forms of flax in late gestation and lactation: Effects on sow and litter performances, endocrinology, and immune response. *J Anim Sci* 2010; 88: 225-237.

## **Illustrations, Figures, Photographs**

Four copies of all figures or photographs should be included with the submitted manuscript. Figures submitted electronically should be in JPEG or TIFF format with a 300 dpi minimum resolution and in grayscale or CMYK (not RGB). Printed submissions should be on high-contrast glossy paper, and must be unmounted and untrimmed, with a preferred size between 4 x 5 inches and 5 x 7 inches (10 x 13 cm and 13 x 18 cm). The figure number, name of first author and an arrow indicating "top" should be typed on a gummed label and affixed to the back of each illustration. If arrows are used these should appear in a different color to the background color. Titles and detailed explanations belong in the legends, which should be submitted on a separate sheet, and not on the illustrations themselves. Written informed consent for publication must accompany any photograph in which the subject can be identified. Written copyright permission, from the publishers, must accompany any illustration that has been previously published. Photographs will be accepted at the discretion of the Editorial Board.