

# Silibinin improves the cytotoxicity of methotrexate in chemo resistant human rhabdomyosarcoma cell lines

Saad A. Hussain, MSc, PhD, Bushra H. Marouf, MSc.

## ABSTRACT

**الأهداف:** دراسة إن كان للسليبينين دور في السيطرة على المقاومة الكيميائية لميتوتريكسات الساركومة العضلية المخططة البشرية.

**الطريقة:** أجريت هذه الدراسة في قسم الأدوية والسموم، كلية الصيدلة، جامعة بغداد، بغداد، العراق خلال الفترة من أكتوبر 2012م حتى مارس 2013م. أجريت دراسة في المختبر لتحريض مقاومة MTX في خلية مثوتركسات المقاومة للساركومة العضلية. المخططة البشرية وتم علاج الخلايا بتراكيز مختلفة من MTX، أو السليبين لوحده، أو كلاهما. قمنا بتحديد حياة الخلية باستخدام مقياس MTT.

**النتائج:** يساعد SDH في طريقة تركيز الخلايا المعتمدة على مقاومة الحساسية في خلايا MTX للوصول إلى أعلى تركيز سام لخلايا MTX، ويقلل من IC50 في MTX حوالي 17.8 طية. كما ارتبط انخفاض IC50 لخلايا MTX بشكل عكسي مع ارتفاع تركيز SDH (R=0.78, p=0.04).

**خاتمة:** يحسن SDH من حساسية ميتوتريكسات الساركومة العضلية المخططة البشرية للنشاط السمي لخلايا MTX في طريقة تركيز الخلايا المعتمدة.

**Objectives:** To investigate whether silibinin (SDH) could overcome chemoresistance of methotrexate (MTX)-resistant human rhabdomyosarcoma (hRD).

**Methods:** This study was conducted at the Department of Pharmacology and Toxicology, College of Pharmacy, University of Baghdad, Baghdad, Iraq from October 2012 to March 2013. In this in vitro study, resistance to MTX was induced in hRD cell line, the cells were treated with different concentrations of MTX or SDH alone, and in combination. Cell viability was determined by tetrazolium assay.

**Results:** The SDH in a concentration-dependent pattern, enhanced the sensitivity of MTX-resistant cells to the maximum cytotoxic concentration of

MTX, and decreased the IC<sub>50</sub> (concentration resulting in 50% inhibition of cell growth) of MTX by 17.8 fold. The decrease in IC<sub>50</sub> of MTX was negatively correlated with increasing SDH concentrations with R<sup>2</sup> = 0.78 and p=0.04.

**Conclusion:** The SDH improves the sensitivity of MTX-resistant hRD cell lines to the cytotoxic activity of MTX in concentration-dependent pattern.

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*From the Department of Pharmacology and Toxicology (Hussain), College of Pharmacy, University of Baghdad, Baghdad, and the Department of Pharmacology and Toxicology (Marouf), School of Pharmacy, Faculty of Medical Sciences, University of Sulaimani, Kurdistan, Iraq.*

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*Address correspondence and reprint request to: Professor Saad A. Hussain, Department of Pharmacology and Toxicology, College of Pharmacy, University of Baghdad, Baghdad, Iraq. Tel. +964 (790) 1712624. E-mail: saad\_alzaidi@yahoo.com*

Drug resistance is one of the main causes of treatment failure and mortality in cancer patients. Drug resistance concomitant with an invasion has been found in many cancer cells isolated from patients with different types of tumors,<sup>1</sup> that cells have these properties may be due to an increase in metastatic potential following chemotherapeutic insults.<sup>2</sup> Thus, there is an urgent need for novel treatment strategies to overcome drug resistance and tumor metastasis. Methotrexate (MTX) is used, mostly in combination with other cytotoxic agents, for the treatment of many neoplasms including acute leukemia and certain solid tumors,<sup>3</sup> it is also used in non-neoplastic disorders, and as an anti-inflammatory

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and/or immunosuppressive drug.<sup>4,5</sup> After transport into the cytoplasmic milieu through the reduced folate carrier, MTX was polyglutamated through an energy consuming reaction catalyzed by folylpolyglutamate synthase (FPGS).<sup>6</sup> Resistance to the effect of MTX was reported after prolonged treatment with consequent attenuation of its efficacy. However, combination treatment of MTX with other drugs that could modulate the expression of genes involved in MTX resistance, and be an adequate strategy to prevent the development of resistance.<sup>7</sup> Many molecular mechanisms are suggested for MTX resistance, including decreased cellular uptake via reduced folate carriers (RFC).<sup>8</sup> The increase of MTX extrusion to the extracellular compartments was mainly attributed to the excessive expression many membrane-bound multi-drug resistance proteins, especially MRP1-4 group that contributed to a specific pattern of MTX resistance.<sup>9</sup> Treatment failure and remarkable toxicity of chemotherapeutic agents encouraged the search for many alternative approaches like the use of phytochemicals, which are thought to be effective with a wide margin of safety. Moreover, many current advances in drug development have revealed cancer preventive and curative efficacies of many phytochemicals.<sup>10</sup> Silibinin di-hemisuccinate (SDH), a flavonoid antioxidant from milk thistle (*Silybum marianum* L.), has been used extensively for many years for the treatment of liver disorders.<sup>11</sup> It recently received attention due to its chemopreventive and anticancer activity.<sup>12</sup> In vitro and in vivo studies have shown that SDH is a potent sensitizer for apoptosis induced by a range of anticancer drugs.<sup>13,14</sup> Additionally, several in vitro and in vivo combination studies of SDH and chemotherapeutic drugs were carried out to analyze the effects of such a combination on growth inhibition, cell cycle regulation, and apoptosis.<sup>15-17</sup> However, no report was found during the literature search for the involvement of SDH in sensitizing chemoresistant malignant cells to MTX. In this study, we evaluated the effect of SDH in increasing sensitivity of resistant human rhabdomyosarcoma (hRD) cells to MTX, and the dose response relationship for this effect.

**Methods.** This study was conducted at the Department of Pharmacology and Toxicology, College of Pharmacy, University of Baghdad, Baghdad, Iraq from October 2012 to March 2013. Methotrexate (Lederle, Hampshire, UK) and SDH (Madaus, Koeln, Germany) stock solutions were obtained as pharmaceutical preparations at concentrations 2 mM (MTX) and 1 mM (SDH). These drugs were stored at 4°C. Immediately before use, they were diluted with

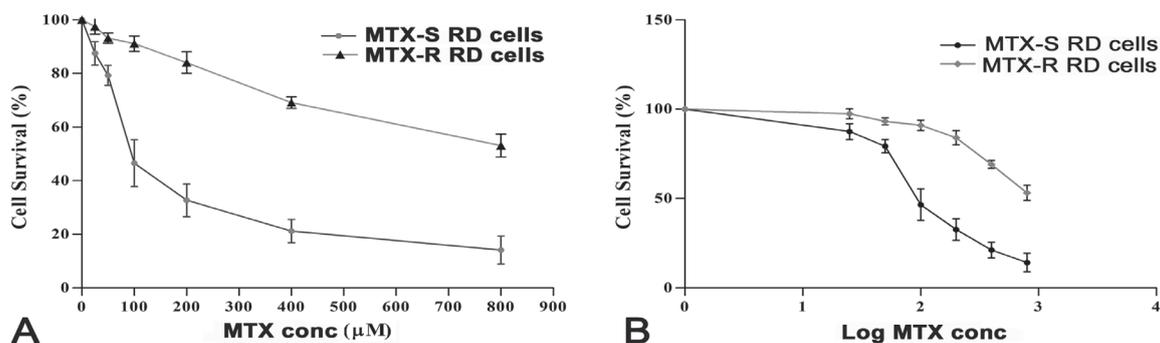
RPMI 1640 prepared as for cell culture. The tetrazolium (MTT) (Sigma Chemical Co, Poole, Dorset, UK) was dissolved in phosphate buffered saline (PBS) to produce a stock solution of 5 mg/ml, which was stored at 4°C. The local scientific committee approved the research protocol. This study used human hRD line as previously described.<sup>18</sup> The hRD cell line was obtained from the Research Center of Biotechnology, Al-Nahrain University, Baghdad, Iraq. The hRD cell lines were maintained in RPMI 1640 (Sigma Chemical Co, Poole, Dorset, UK) supplemented with 10% fetal bovine serum (Life Technologies, Inc., Scotland, UK), 2 mM L-glutamine (Sigma Chemical Co, Poole, Dorset, UK), 100 U/ml penicillin, and 100 µg/ml streptomycin (Invitrogen, Carlsbad, CA, USA). Cells were grown as attached monolayers, and were incubated at 37°C in a humidified atmosphere with 5% CO<sub>2</sub>. Resistant hRD cells were obtained in the laboratory through the incubation with stepwise concentrations of MTX as previously described.<sup>19</sup> Cytotoxicity was assessed using the MTT assay as described by Mosmann.<sup>20</sup> Briefly, hRD cells (2x10<sup>5</sup> cells/mL) were plated in 100 µL in each well of 96-well round-bottomed microtiter plates. Methotrexate and SDH (each alone or in combinations), were added at the desired concentrations, and the plates were incubated at 37°C for 72 hours in a 5% CO<sub>2</sub> humidified atmosphere. At the end of the incubation period, 10 µL of a stock solution of 5 mg/mL MTT, were added to each well, and the plates were incubated for another 4 hours at 37°C. Absorbance was measured using an enzyme-linked immunosorbent assay (ELISA) plate reader (LabSystems Multiscan RC, Helsinki, Finland) at a wavelength of 540 nm with reference at 650 nm. Growth inhibition was measured by dividing the mean absorbance of treated wells per mean absorbance of control wells (drug-free wells), and is expressed as a percentage. The inhibitory concentrations of 50% of cells (IC<sub>50</sub>) values were defined as the drug concentrations, at which cell growth was inhibited by 50% compared with drug-free controls. Values were expressed as mean ± standard deviation, the values were statistically evaluated using linear regression test, unpaired student's t-test, and one-way analysis of variance (ANOVA), supported by Bonferroni's post hoc analysis. Values with *p*<0.05 were considered significant. Analysis was performed using Graph Pad Prism software for Windows version 5 (Graph Pad Software Inc, San Diego, CA, USA).

**Results.** Figure 1 shows the effect of different concentrations of MTX on the survival of MTX-sensitive hRD cell line, where maximum effect achieved

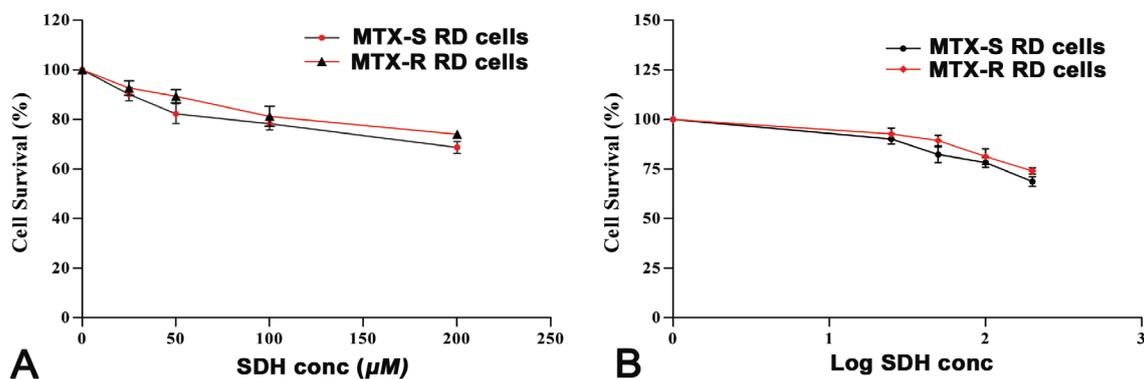
(14.2%) with 800  $\mu\text{M}$  MTX. Meanwhile, in MTX-resistant hRD cells, the same concentration of MTX (800  $\mu\text{M}$ ) decreases cell survival only to 53.1%, and the  $\text{IC}_{50}$  for MTX was significantly increased ( $p < 0.05$ ) from 102.3  $\mu\text{M}$  to 1258  $\mu\text{M}$  (12.3 fold) (Table 1). In Figure 2, the maximum concentration of SDH (200  $\mu\text{M}$ ) decreases the MTX-sensitive cell survival to 68.7%, while in MTX-resistant cell line, this concentration of SDH decreased survival to 74%; the  $\text{IC}_{50}$  for SDH was significantly increased ( $p < 0.05$ ) by 5-fold, as shown in Table 1. Figure 3 shows that incubation of MTX-resistant hRD cells with 800  $\mu\text{M}$  MTX and increasing concentrations of SDH (0, 25, 50, 100 and 200  $\mu\text{M}$ ) improves the cytotoxic effect of the given concentration of MTX and produces dramatic decrease in cell survival, achieving only 2.4% survival with maximum concentration of SDH (200  $\mu\text{M}$ ). Table 2 clearly shows that SDH decreases the  $\text{IC}_{50}$  for MTX in a concentration-dependent pattern, and the maximum

SDH concentration significantly decreases ( $p < 0.05$ ) the  $\text{IC}_{50}$  for MTX to 17.8-fold compared to its value when used alone in MTX-resistant hRD cells. The decrease in  $\text{IC}_{50}$  of MTX was negatively correlated with increasing SDH concentrations, with  $R^2 = 0.78$  and  $p = 0.04$  (Figure 4).

**Discussion.** Although many efforts are carried out to improve chemotherapeutic response in cancer cells, development of drug resistance remains a major challenge. In fact, only 50-60% of cancers are responsive to chemotherapy,<sup>21</sup> indicating that the outcome is still far from optimum. The use of alternative anti-cancer agents, particularly those obtained from natural sources, may be a good choice in this regard.<sup>22</sup> Silibinin is a natural chemical that shows promising anti-cancer activity in several studies, including a phase I clinical trials.<sup>23,24</sup> In vitro studies confirm that SDH at a range of 25-100  $\mu\text{mol/L}$  inhibits cell viability, likely through



**Figure 1** - Effects of different doses of methotrexate (MTX) on the survival of MTX-S (sensitive) and MTX-R (resistant) human rhabdomyosarcoma cell (RD) line expressed in ordinary (A), and log (B) concentration scale. SDH - silibinin



**Figure 2** - Effects of different doses of silibinin on the survival of methotrexate (MTX)-sensitive (S) and MTX-resistant (R) human rhabdomyosarcoma (RD) cell line, expressed in ordinary (A), and log (B) concentration (conc) scale. SDH - silibinin

**Table 1** - Sensitivity of MTX-sensitive and MTX-resistant hRD cell line to the effects of MTX and SDH (measured as IC<sub>50</sub>).

Type of treatment	n	IC <sub>50</sub> (μM)	
		MTX-sensitive hRD cells	MTX-resistant hRD cells
MTX	6	102.3 ± 13.4	1258 ± 168.1*
SDH	6	15848 ± 910.4	79432 ± 680.2*

IC<sub>50</sub> - concentration resulting in 50% inhibition of cell growth, hRD - rhabdomyosarcoma; MTX - methotrexate, SDH - silibinin, \*significantly different compared with MTX-sensitive hRD cells (*p*<0.05)

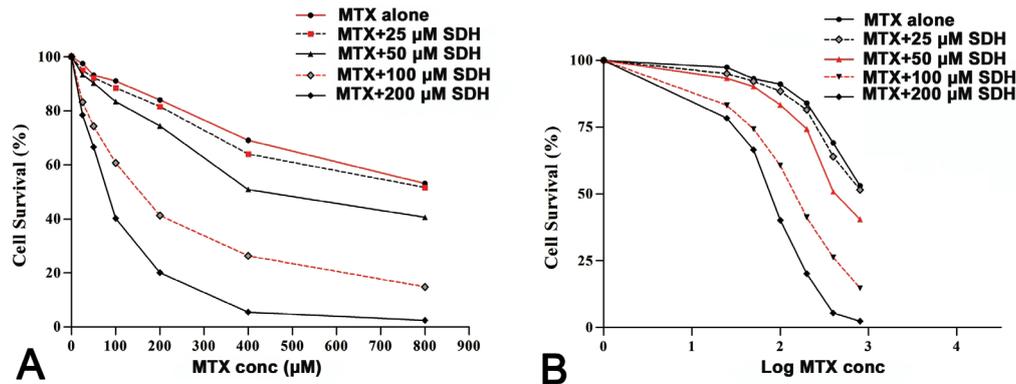
**Table 2** - Effect of different concentrations of silibinin dihemisuccinate (SDH) on the sensitivity of MTX-resistant hRD cells to 800 μM MTX (measured as IC<sub>50</sub>).

Silibinin (μM)	Methotrexate IC <sub>50</sub> (μM)
0	1258 ± 168.1 <sup>a</sup>
25	1122 ± 86.7 <sup>a</sup>
50	562.3 ± 62.3 <sup>b</sup>
100	158.5 ± 15.9 <sup>c</sup>
200	70.8 ± 10.1 <sup>d</sup>

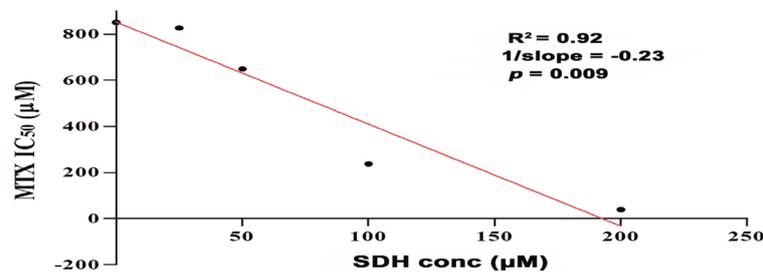
IC<sub>50</sub> - concentration resulting in 50% inhibition of cell growth, values with non-identical superscripts (a, b, c, d) are significantly different (*p*<0.05)

cell cycle arrest and induction of apoptosis.<sup>25,26</sup> While SDH has been evaluated in many types of tumors, very little is known regarding its effects in MTX-resistant hRD.

In the present study, we provide evidence that SDH improves the response to the cytotoxic effects in MTX-resistant hRD cells. There is a well-documented cross-resistance between MTX and SDH in MTX-resistant hRD cells. The IC<sub>50</sub> of SDH in our model was 1258 ± 168.1 μM, and cytotoxicity was confirmed by MTT assay. These results are consistent with previous studies regarding the effect of the other polyphenol, the quercetin, in osteosarcoma cells.<sup>27</sup> In the present study, hRD cells were treated with different concentrations of MTX for 72 hand cell viability was measured by the MTT assay. As shown in Figure 2 and Table 1, hRD cells were resistant to MTX in that the IC<sub>50</sub> of MTX was 12.3-fold higher in these cells compared to their parent cell line. To investigate whether SDH was effective in improving sensitivity of resistant hRD cells to MTX, the cytotoxicity of combination of MTX with different concentrations of SDH in MTX-resistant hRD cells



**Figure 3** - Effects of different doses of silibinin on the survival of methotrexate (MTX)-resistant human rhabdomyosarcoma cells incubated with different concentrations of MTX, expressed in ordinary (A), and log (B) MTX concentration scale. SDH - silibinin



**Figure 4** - Correlation between silibinin concentration and methotrexate (MTX) IC<sub>50</sub> required to inhibit growth of human rhabdomyosarcoma cell line. IC<sub>50</sub> - concentration resulting in 50% inhibition of cell growth, SDH - silibinin

was determined. The results showed that MTX/SDH combination produced a dramatic reversal of MTX resistance in hRD cells. As shown in Table 2, the IC<sub>50</sub> for MTX decreased with increasing concentrations of SDH up to 200 μM. Silibinin reduced the IC<sub>50</sub> of MTX, in a dose-dependent pattern from 1258 μM to 70.8 μM (decrease IC<sub>50</sub> up to 17.8-fold). In addition to its well-known cytoprotective activity, SDH enhances the activity of chemotherapeutic agents.<sup>28</sup> In this respect, many researchers reported that SDH effectively increases the sensitivity of prostate cancer cells to the cytotoxicity induced by many chemotherapeutic agents, including doxorubicin, cisplatin, carboplatin, and mitoxantrone.<sup>15-17</sup> However, bioavailability of SDH was considered as a major problem that limits its therapeutic utility, it can be attributed to its large polyphenolic structure and low water solubility.<sup>29</sup> Accordingly, modified formulations have tried to solve this problem.<sup>30</sup> In many MTX-responsive cancers, the currently adopted treatment programs are based on a combination of MTX with other agents, including doxorubicin and cisplatin, where MTX is the most active one.<sup>31</sup> Despite the improvement of response, a considerable number of patients develop MTX resistance and die due to disease progression.<sup>21</sup> Accordingly, additional therapeutic agents need to be evaluated to improve the survival of MTX-resistant patients.

The use of sensitizers is one of the available strategies to reverse resistance to chemotherapy. Silibinin has been demonstrated to be an effective chemopreventive and chemotherapeutic agent,<sup>32</sup> without any toxic or adverse effects.<sup>33,34</sup> Therefore, it may have potential clinical application in combination chemotherapy. Although the present study did not reveal the exact mechanisms, through which SDH reverses chemoresistance, our results demonstrate that SDH produced a dramatic reversal of MTX resistance and enhanced MTX-induced cytotoxicity in hRD cells resistant to MTX. This finding is in agreement with that reported by Zhou et al,<sup>35</sup> where SDH significantly restores the sensitivity of chemoresistant human ovarian carcinoma cells to paclitaxel. Many mechanisms are suggested for the antiproliferative activity of SDH, including the inhibition of the Cdk4 pathway, increase in p53 expression, down-regulation of surviving and cyclin D1, phosphorylation of ERK1/2 and nuclear phospho-p65, cleavage of caspases, and mitochondrial release of cytochrome c, and apoptosis-inducing factors.<sup>36-39</sup> Additionally, MTX is known to be a P-glycoprotein (P-gp) substrate, and emergence of resistance may be

due to over expression of this membrane protein;<sup>40</sup> the inhibition of P-gp by SDH may lead to increased accumulation of MTX with a consequent increase in sensitivity of resistant cells to the cytotoxic activity of MTX.<sup>41</sup> Further studies are required to investigate the exact molecular mechanisms behind the effect of SDH in this regard.

In conclusion, SDH improves the sensitivity of MTX-resistant hRD cell lines to the cytotoxic activity of MTX in concentration-dependent pattern.

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## References

- Weinstein RS, Jakate SM, Dominguez JM. Relationship of the expression of the multidrug resistance gene product (P-glycoprotein) in human colon carcinoma to local tumor aggressiveness and lymph node metastasis. *Cancer Res* 1991; 51: 2720-2726.
- De Larco JE, Wuertz BR, Manivel JC. Progression and enhancement of metastatic potential after exposure of tumor cells to chemotherapeutic agents. *Cancer Res* 2001; 61: 2857-2861.
- Bertino JR. Karnofsky memorial lecture: ode to methotrexate. *J Clin Oncol* 1993; 11: 5-14.
- Cronstein BN. Molecular therapeutics: methotrexate and its mechanism of action. *Arthritis Rheum* 1996; 39: 1951-1960.
- Fabre G, Fabre I, Matherly LH, Cano JP, Goldman ID. Synthesis and properties of 7-hydroxymethotrexate polyglutamyl derivatives in Ehrlich ascites tumor cells in vitro. *J Biol Chem* 1984; 259: 5066-5072.
- Fabre G, Matherly LH, Favre R, Catalin J, Cano JP. In vitro formation of polyglutamyl derivatives of methotrexate and 7-hydroxymethotrexate in human lymphoblastic leukemia cells. *Cancer Res* 1983; 43: 4648-4652.
- Rots MG, Pieters R, Kaspers GJ, Veerman AJ, Peters GJ, Jansen G. Classification of ex vivo methotrexate resistance in acute lymphoblastic and myeloid leukemia. *Br J Haematol* 2000; 110: 791-800.
- Sirotnak FM, Tolner B. Carrier-mediated membrane transport of folates in mammalian cells. *Annu Rev Nutr* 1999; 19: 91-122.
- Stark M, Rothem L, Jansen G, Scheffer GL, Goldman ID, Assaraf YG. Antifolate resistance associated with loss of MRP1 expression and function in Chinese hamster ovary cells with markedly impaired export of folate and cholate. *Mol Pharmacol* 2003; 64: 220-227.
- Surh YJ. Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer* 2003; 3: 768-780.
- Ferenci P, Dragosics B, Dittrich H, Frank H, Benda L, Lochs H, et al. Randomized controlled trial of silymarin treatment in patients with cirrhosis of the liver. *J Hepatol* 1989; 9: 105-113.
- Varghese L, Agarwal C, Tyagi A. Silibinin efficacy against human hepatocellular carcinoma. *Clin Cancer Res* 2005; 11: 8441-8448.

13. Deep G, Singh RP, Agarwal C, Kroll DJ, Agarwal R. Silymarin and silibinin cause G1 and G2-M cell cycle arrest via distinct circuitries in human prostate cancer PC3 cells: a comparison of flavanone silibinin with flavanone mixture silymarin. *Oncogene* 2006; 25: 1053-1069.
14. Tyagi AK, Agarwal C, Chan DC, Agarwal R. Synergistic anti-cancer effects of silibinin with conventional cytotoxic agents doxorubicin, cisplatin and carboplatin against human breast carcinoma MCF-7 and MDA-MB468 cells. *Oncol Rep* 2004; 11: 493-499.
15. Dhanalakshmi S, Agarwal P, Glode LM, Agarwal R. Silibinin sensitizes human prostate carcinoma DU145 cells to cisplatin and carboplatin-induced growth inhibition and apoptotic death. *Int J Cancer* 2003; 106: 699-705.
16. Tyagi AK, Singh RP, Agarwal C, Chan DC, Agarwal R. Silibinin strongly synergizes human prostate carcinoma DU145 cells to doxorubicin-induced growth inhibition, G2-M arrest, and apoptosis. *Clin Cancer Res* 2002; 8: 3512-3519.
17. Flaig TW, Su LJ, Harrison G, Agarwal R, Glode LM. Silibinin synergizes with mitoxantrone to inhibit cell growth and induce apoptosis in human prostate cancer cells. *Int J Cancer* 2007; 120: 2028-2033.
18. Cocker HA, Pinkerton CR, Kelland LR. Characterization and modulation of drug resistance of human pediatric rhabdomyosarcoma cell lines. *Br J Cancer* 2000; 83: 338-345.
19. Selga E, Noe V, Ciudad CJ. Transcriptional regulation of aldo-ketoreductase 1C1 in HT29 human colon cancer cells resistant to methotrexate: role in the cell cycle and apoptosis. *Biochem Pharmacol* 2008; 75: 414-426.
20. Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods* 1983; 65: 55-63.
21. Lewis IJ, Nooij MA, Whelan J, Sydes MR, Grimer R, Hogendoorn PC, et al. Improvement in histologic response but not survival in osteosarcoma patients treated with intensified chemotherapy: a randomized phase III trial of the European Osteosarcoma Intergroup. *J Natl Cancer Inst* 2007; 99: 112-128.
22. Kong JM, Goh NK, Chia LS, Chia TF. Recent advances in traditional plant drugs and orchids. *Acta Pharmacol Sin* 2003; 24: 7-21.
23. Lah JJ, Cui W, Hu KQ. Effects and mechanisms of Silibinin on human hepatoma cell lines. *World J Gastroenterol* 2007; 13: 5299-5305.
24. Hoh C, Boocock D, Marczylo T, Singh R, Berry DP, Dennison AR, et al. Pilot study of oral silibinin, a putative chemopreventive agent, in colorectal cancer patients: silibinin levels in plasma, colorectum, and liver and their pharmacodynamic consequences. *Clin Cancer Res* 2006; 12: 2944-2950.
25. Kauntz H, Bousserouel S, Gosse F, Raul F. Silibinin triggers apoptotic signaling pathways and autophagic survival response in human colon adenocarcinoma cells and their derived metastatic cells. *Apoptosis* 2011; 16: 1042-1053.
26. Tyagi A, Agarwal C, Harrison G, Glode LM, Agarwal R. Silibinin causes cell cycle arrest and apoptosis in human bladder transitional cell carcinoma cells by regulating CDK1-CDK-cyclin cascade, and caspase 3 and PARP cleavages. *Carcinogenesis* 2004; 25: 1711-1720.
27. Xie X, Yin J, Jia Q, Wang J, Zou C, Brewer KJ, et al. Quercetin induces apoptosis in the methotrexate-resistant osteosarcoma cell line U2-OS/MTX300 via mitochondrial dysfunction and dephosphorylation of Akt. *Oncol Rep* 2011; 26: 687-693.
28. Deep G, Agarwal R. New combination therapies with cell-cycle agents. *Curr Opin Invest Drugs* 2008; 9: 591-604.
29. Gazak R, Svobodova A, Psotova J, Sedmera P, Prikrylova V, Walterova D, et al. Oxidised derivatives of Silybin and their antiradical and antioxidant activity. *Bioorg Med Chem* 2004; 12: 5677-5687.
30. Deep G, Agarwal R. Antimetastatic efficacy of silibinin: molecular mechanisms and therapeutic potential against cancer. *Cancer Metastasis Rev* 2010; 29: 447-463.
31. Delepine N, Delepine G, Bacci G, Rosen G, Desbois JC. Influence of methotrexate dose intensity on outcome of patients with high-grade osteogenic osteosarcoma. Analysis of the literature. *Cancer* 1996; 78: 2127-2135.
32. Singh RP, Agarwal R. Mechanisms and preclinical efficacy of silibinin in preventing skin cancer. *Eur J Cancer* 2005; 41: 1969-1979.
33. Flaig TW, Gustafson DL, Su LJ, Zirrolli JA, Crighton F, Harrison GS. A phase I and pharmacokinetic study of silybinphytosome in prostate cancer patients. *Invest New Drugs* 2007; 25: 139-146.
34. Mulrow C, Lawrence V, Jacobs B, Dennehy C, Sapp J, Ramirez G, et al. Milk thistle: effects on liver disease and cirrhosis and clinical adverse effects. *Evid Rep Technol Assess* 2000; 21: 1-3.
35. Zhou L, Liu P, Chen B, Wang Y, Wang X, Internati MC, et al. Silibinin restores paclitaxel sensitivity to paclitaxel-resistant human ovarian carcinoma cells. *Anticancer Res* 2008; 28: 1119-1127.
36. Karim BO, Rhee KJ, Liu G, Zheng D, Huso DL. Chemoprevention utility of silibinin and Cdk4 pathway inhibition in Apc-<sup>-/+</sup> mice. *BMC Cancer* 2013; 13: 157-167.
37. Tyagi A, Raina K, Singh RP, Gu M, Agarwal C, Harrison G, et al. Chemopreventive effects of silymarin and silibinin on N-butyl-N-(4-hydroxybutyl) nitrosamine induced urinary bladder carcinogenesis in male ICR mice. *Mol Cancer Ther* 2007; 6: 3248-3255.
38. Singh RP, Tyagi A, Sharma G, Mohan S, Agarwal R. Oral silibinin inhibits in vivo human bladder tumor xenograft growth involving down-regulation of survivin. *Clin Cancer Res* 2008; 14: 300-308.
39. Zeng J, Sun Y, Wu K, Li L, Zhang G, Yang Z, et al. Chemopreventive and chemotherapeutic effects of intravesical silibinin against bladder cancer by acting on mitochondria. *Mol Cancer Ther* 2011; 10: 104-116.
40. Breedveld P, Beijnen JH, Schellens JH. Use of P-glycoprotein and BCRP inhibitors to improve oral bioavailability and CNS penetration of anticancer drugs. *Trends Pharmacol Sci* 2006; 27: 17-24.
41. Bansal T, Jaggi M, Khar RK, Talegaonkar S. Emerging significance of flavonoids as P-glycoprotein inhibitors in cancer chemotherapy. *J Pharm Pharm Sci* 2009; 12: 46-78.